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의학박사 학위논문

저산소 유도인자 표적항암제
Diacetoxyscirpenol의 발굴 및 전임상
평가에 관한 연구

Discovery and preclinical evaluation of
diacetoxyscirpenol as a HIF-targeting
anticancer agent

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ABSTRACT

Hypoxia activates hypoxia-inducible factor 1, which promotes the progression of malignancy by stimulating angiogenesis and by augmenting the ability of tumors to survive. Thus, HIF-1 is one of the most compelling targets for treating cancers. The aim of this study was to find a small molecule that inhibits HIF-1 under hypoxia in cancer cells. 7,280 compounds in a chemical library were tested in a cancer cell line expressing luciferase HIF-dependently. Through three rounds of screening, I finally picked up a compound that originates from a marine bacterium parasitizing red alga. The antibiotic potently inhibited HIF-1 expression and its transcriptional activity in cancer cells exposed to hypoxia. Through two-step fractionation, diacetoxyscirpenol was purified and identified as a HIF-inhibiting ingredient. Mechanistically, diacetoxyscirpenol inhibits the synthesis of HIF-1 α protein and also interferes with the dimerization of HIF-1 α and ARNT. It attenuates HIF-mediated gene expression in cancer cells exposed to hypoxia, and by doing so reduces tumorigenic and angiogenic potentials of cancer cells. More importantly, diacetoxyscirpenol retarded tumor growth in mice, and reduced HIF-1 α expression and vascular formation in the tumors. Overall, diacetoxyscirpenol is considered a potential drug deregulating the HIF-1

signaling pathway, and it could be beneficially employed for treating malignant tumors with hypoxic microenvironment.

Keywords : Cancer, Hypoxia, HIF-1, Drug screening, Diacetoxyscirpenol

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LIST OF ABBREVIATIONS

HIF : Hypoxia-inducible factor

HRE : Hypoxia response element

ARNT : Aryl hydrocarbon receptor nuclear translocator

PAS : PER-ARNT-SIM

PHD : Prolyl hydroxylase

FIH : Factor inhibiting HIF

CBP : CREB binding protein

pVHL : Von hippel-lindau protein

DMOG : Dimethyloxaloylglycine

IRES : Internal ribosome entry site

RCC : Renal cell carcinoma

DAS : Diacetoxyscirpenol

IC : Inhibitory concentration

EPO : Erythropoietin

LOX : Lysyl oxidase

CA9 : Carbonic anhydrase 9

CITED2 : Cbp/p300 interacting transactivator

VEGF : Vascular endothelial growth factor

HUVEC : Human umbilical vein endothelial cells

DAPI : 4'6-diamidino-2-phenylindole

FBS : Fetal bovine serum

SD : Standard deviation

INTRODUCTION

Hypoxia is a common feature in solid tumors and plays crucial roles for tumor progression and angiogenesis [Harris et al., 2002; Pugh et al., 2003]. To survive under a hypoxic microenvironment, cancer cells require the activation of many genes essential for their adaptation to the condition, which is mainly driven by the transcription factor HIF-1 (hypoxia-inducible factor 1) [Wang et al., 1995]. The HIF family are composed of α and β subunits [Pugh et al., 1997; Bardos et al., 2005]. There are well known isoform of HIF- α : HIF-1 α , HIF-2 α (alternatively named endothelial PAS domain protein-1, EPAS-1). HIF-1 β also known as aryl hydrocarbon receptor nuclear translocator (ARNT) is a common binding partner for HIF-1 α and HIF-2 α [Rocha et al., 2007]. The HIF subunits share basic helix-loop-helix (bHLH) and Per/ARNT/Sim (PAS) domains [Semenza et al., 1985]. In addition, they possess an oxygen-dependent degradation domain (ODDD), which renders HIF- α protein labile in the presence of oxygen. HIF-1 α and HIF-2 α contain two transactivation domains (TADs), namely, the N-terminal TAD (NAD) and the C-terminal TAD (CAD). HIF-1 α is rapidly degraded under normoxic conditions, when it is hydroxylated at two proline residues (Pro 402 and Pro 564) within the ODDD by hydroxylase

domain (PHD) 1-3 [Fandrey et al., 2006]. This post-translational modification serves as a recognition motif for subsequent ubiquitination by the E3 ubiquitin ligase activity of the von Hippel-Lindau tumor (pVHL) suppressor protein, thus targeting HIF-1 α for degradation by the 26S proteasome [Ivan et al., 2001; Jaakkola et al., 2001; Maxwell et al., 1999]. In hypoxic conditions, hydroxylation of HIF-1 α does not occur, and the α subunit is free to migrate to the nucleus, bind to its counterpart, and thus act as a transcription factor along with other cofactors. An additional level of regulation mediated by cellular oxygen levels is given by the factor inhibiting HIF (FIH-1) [Dayan et al., 2006], an asparagine hydroxylase that hydroxylates the N803 residue in the carboxyl-terminal transactivation domain (CAD) of HIF-1 α . In hypoxic conditions, N803 is not hydroxylated by FIH-1; therefore, HIF-1 α can interact with its transcriptional coactivators, p300 and CBP (CREB binding protein), and hence further promote transcription of genes that have hypoxia responsive enhancer elements in their promoter region [Mahon et al., 2001; Lando et al., 2002].

The expression of HIF-1 in tumor specimens positively correlates with tumor aggressiveness and poor prognosis [Birner et al., 2000]. Also, many tumor grafting studies have supported the positive effects of HIF-1 on tumorigenesis, tumor growth, and tumor invasion [Ryan et al., 2000]. Given clinical and experimental evidence, HIF-1 has been believed as a promising target for

treating tumors. Indeed, many types of HIF-1 inhibitors have been discovered, and some of them are currently under clinical trials. HIF-1 inhibitors act through various mode-of-actions as followings (Table 1).

Table 1. HIF-1 Pharmacological inhibitors

Inhibitors	Target	References
Bortezomib	FIH / P300 interaction	Shin et al., 2008
Nutlin-3	FIH / P300 interaction	Lee et al., 2009
Echinomycin	DNA binding	Kong et al., 2005
17AAG	HSP90	Isaacs et al., 2002
2-ME	Microtubule destabilizer	Ricker et al., 2004
YC-1	Soluble guanylyl cycle	Yeo et al., 2003
Topotecan	Topoisomerase I	Rapisarda et al., 2004
Camtothecin	Topoisomerase I	Lou et al., 2010
Digoxin	Not determined	Zhang et al., 2008
Rapamycin	mTOR	Hudson et al., 2002
LY294002	PI3K	Zhong et al., 2000
Chaetocin	HIF-1 α pre-mRNA splicing	Lee et al., 2011

Bortezomib, nutlin-3, and echinomycin functionally inhibit the HIF-1-mediated gene expressions [Shin et al., 2008; Lee et al., 2009; Kong et al., 2005]. HSP90 inhibitors, microtubule disruptors, and YC-1 destabilize HIF-1 α in the post-translational level [Isaacs et al., 2002 Ricker et al., 2004; Yeo et al., 2003].

Several agents, such as topotecan, camptothecin, digoxin, rapamycin, LY294002 and chaetocin, have been reported to block de novo synthesis of HIF-1 α protein [Raspisarda et al., 2004; Lou et al., 2010; Zhang et al., 2008; Hudson et al., 2002; Zhong et al., 2000; Lee et al., 2011].

In spite of many efforts to discover small-molecule inhibitors of the HIF signaling pathway, none of the inhibitors has been identified to directly (physically) target the HIF- α proteins. Actually, all of them indirectly control the HIF signaling pathway by inhibiting the non-selective regulators that also participate in the de novo synthesis or stabilization of proteins other than HIF- α . Nonetheless, some of those inhibitors, such as camptothecin, topotecan, bortezomib, echinomycin, 17-AAG, were FDA-approved for cancer treatment. Yet, they have not been clinically used so far because they could make adverse effects due to lack of specificity. To realize the HIF-targeting cancer therapy, the molecules targeting HIF- α *per se* should be discovered, so it is still worth while to search for new HIF inhibitors.

The aim of this study was to find a small molecule that inhibits HIF-1 specific under hypoxia in cancer cells. A human hepatoma cell line, which stably expresses the luciferase reporter gene targeted by HIF-1, was established and used for searching for HIF-1 inhibitors in a chemical library containing 7,280 compounds. Finally, I picked up a compound that originates from a marine

bacterium living on red alga, and identified this compound as diacetoxyscirpenol.

Moreover, I examined how the compound inhibits HIF-1 and whether it has anti-cancer and anti-angiogenic activities.

MATERIALS AND METHODS

1. Cell culture and transfection

Hep3B (hepatoma), A549 (lung cancer), HCT116 (colon cancer) and LLC1 (mouse lung cancer) cell lines were obtained from the American Type Culture Collection (Manassas, VA). MKN45 (gastric cancer) was obtained from Korean Cell Line Bank (Seoul, Korea). RCC4/VHL^{-/-} (renal cancer) was obtained from European Collection of Authenticated Cell Cultures (London, UK). The cells were cultured in Minimum Essential Media, Dulbecco's Modified Eagle Medium or RPMI-1640, supplemented with 10 % heat-inactivated FBS. Cells were incubated in a humidified atmosphere at 37 °C under 20 % O₂/5 % CO₂ for normoxia or 1 % O₂/5 % CO₂ hypoxia. For transient transfection, cells at about 40 % confluence were transfected with plasmids using Lipofectamin 2000 (Life Technologies, Carlsbad, CA). Cells were allowed to stabilize for 48 hours before being used in experiments.

2. Reagents and antibodies

Diacetoxyscirpenol was purchased from LKT laboratories (Saint Paul, MN), and 17AAG and DMOG from Sigma-Aldrich Corp (St Louis, MO). Anti-HIF-2 α

(#7096) antibody was purchased from Cell Signaling Technology (Danvers, MA), and antibodies against ARNT (sc-8076), β -tubulin (sc-9104) and lamin-B (sc-6216) from Santa Cruz Biotechnology (Santa Cruz, CA). Anti-HIF-1 α antiserum was generated in rabbits against a bacterially expressed fragment encompassing amino acids 418 to 698 of human HIF-1 α .

3. Chemical library and fractionation of marine bacterial metabolites

A chemical library containing 7,280 compounds was kindly provided by Korea Chemical Bank (Daejeon, Korea; <http://www.chembank.org/>). A bacterial strain *Bacillus licheniformis* was isolated from the marine red alga *Gelidium pacificum* collected at Geomun Island in South Korea, and identified based on 18S rRNA analyses (performed in SolGent Co., Ltd., Daejeon, Korea). The bacterium was cultured in 20 L of culture medium consisting of soytone (0.1 %), soluble starch (1.0 %), and seawater (100 %). The cultures were incubated at 29 °C for 20 days on the static condition. The whole culture was filtered through cheesecloth to yield broth and mycelium residue, and the resulted broth were extracted with EtOAc to afford broth extract (1.1 g). The broth extract was subjected to silica gel flash column chromatography. Elution was performed with n-hexane-EtOAc (stepwise, 0 - 100% EtOAc) to yield eight fractions. Fraction 5 (15.5 mg) displayed HIF activity, and was further purified by silica gel flash column

chromatography using n-hexane-EtOAc (stepwise, 10 - 100% EtOAc) to give crude diacetoxyscirpenol (8.0 mg), which was finally purified by LH-20 column chromatography (MeOH) to furnish diacetoxyscirpenol (1.5 mg).

4. Reporter assays

To construct the EPO enhancer-driven luciferase reporter gene, the human EPO enhancer region (5-GGTACCGGCCCTACGTGCTGTCTCACACAGCCTGTCTGACCTCTCGACCTACCGGCCAGATCT-3) was inserted into the multi-cloning site of the pGL4 vector (Promega, Madison, WI). Mutated EPO luciferase was generated by altering the DNA sequence targeted by HIF-1 (CGTG to AAAA). Hep3B cells were transfected with 1 mg of the reporter plasmid using Lipofectamin 2000 (Life Technologies). After stabilized for 48 hours, the Hep3B cell lines stably harboring the reporter plasmid were selected under G418, and five stable cell lines were mixed to rule out the artifact generated by the integration of the plasmid into chromosomes. The cell line was pre-treated with each compound in a chemical library for 4 hours, and then incubated under normoxia or hypoxia for 16 hours. Cells were lysed and luciferase activity was measured using a LB960 luminometer (Berthold Technologies, Oak Ridge, TN). To evaluate the capdependent and the IRES-dependent translation of HIF-1 α mRNA, I used thymidine kinase (TK)

promoter / HIF-1 α 5'-UTR / luciferase and CMV promoter / GFP / HIF-1 α 5'-UTR/luciferase reporters, respectively [Cho et al., 2009]. The β -gal expression plasmid was co-transfected into cells to normalize transfection efficiency.

5. Cell viability assays

An MTT labeling reagent (Sigma-Aldrich, St Louis, MI) was used to evaluate cytotoxicity. Cells were plated in 24-well dishes and cultured in 0.5 ml of medium per well. After cells were treated with each compound for 24 hours, cells were incubated with 5 mg/mL of MTT for 3 hours. Blue formazan crystals were solubilized with acidified isopropanol, and formazan levels were determined at 570 nm using a spectrophotometer.

6. Immunoblotting and immunoprecipitation

Total proteins were separated on an SDS polyacrylamide gel and transferred to Immobilon-P membranes (Millipore, Bedford, MA). Membranes were blocked with 5 % nonfat milk in a 0.1 % Tween-20 / Trisbuffered saline (TTBS) at room temperature for 1 hour, and sequentially incubated overnight at 4 °C with a primary antibody (1:1000) and with horseradish peroxidaseconjugated secondary antibody (1:5000) for 1 hour. The immune complexes were visualized using an Enhanced Chemiluminescence Plus kit (Amersham Biosciences, Piscataway, NJ).

The intensities of HIF-1/2 α blots were quantified using Image J program v1.31 (NIH, Bethesda, MD) and normalized to β -tubulin levels. Data from 3 independent experiments were compared statistically. To analyze the interaction between HIF-1 α and ARNT, cell lysates were incubated with anti-HIF-1 α antibody for 4 hours, and further incubated with protein A/G-Sepharose beads (GE Healthcare BioSciences, Westborough, MA) at 4 °C for 2 hours. Co-precipitated ARNT was identified by immunoblotting with anti-ARNT antibody.

7. Immunofluorescence

A549 cells were cultured on glass coverslips. The cells were fixed with 4 % paraformaldehyde for 30 minutes, washed twice with PBS, blocked with 5 % FBS, and incubated with anti-HIF-1 α antibody for 16 hours. The cells were incubated with anti-rabbit IgG antibody conjugated with Alexa Fluor® 488 for 1 hour, and their nuclei were stained with 30 μ M DAPI for 5 minutes. Finally, the cells were mounted with the Fluorescence Mounting Medium (Thermo Fisher Scientific, Waltham, MA) and photographed under a fluorescence microscope.

8. Secreted VEGF assay

A549 cell was treated with a compound under normoxia and hypoxia for 24

hours. VEGF released from cells was analyzed in medium using the human VEGF ELISA kit provided by Life Technologies. The VEGF levels were calculated based on the standard curve using a recombinant human VEGF 165. Each assay was triplicate to reduce the intra-assay error.

9. Quantitative RT-PCR

Total RNAs were reverse-transcribed, and realtime polymerase chain reaction analyses were performed in 96-well optical plates. Data were analyzed using CFX Manager Software (Bio-Rad Laboratories, Hercules, CA), and values were normalized to β -actin. The sequences (5' to 3') of primers are ACCAAGGGACATCAGATTTTC & GTAGCCATAGTCACAGGATG for lysyl oxidase (LOX); GGAAGAAAACAGTGCCTATG & GTG TAGTCAGAG ACCCCTCA for carbonic anhydrase IX (CA9); ACCCACCTCCCTTATGTAGT & CCAACTAATGCAATTT TTCC for CBP/p300-interacting transactivator with G/D-rich carboxy-terminal domain 2 (CITED2); GGAGGAGAACAAACAGATCA & GTAGGACAGGAAGTTGTTGG for cyclin D1; GTTGCCACTTCCACATAATG & TCATCTGTGCTTTCATGTCA for HIF-1 α ; GTGGACATCTTCCAGGAGTA & GTGATGTTGGACTCCTCAGT for vascular endothelial growth factor (VEGF); and GGACCTGACTGACTACCTCA & AGCTTCTCCTTAATGTCACG for β -actin.

10. Anchorage-independent tumor growth assay

The top agar mixture (RPMI-1640 with 10 % fetal bovine serum, 0.4 % agar, and 2×10^3 A549 cells) was poured into a dish which had been coated with 0.8 % agar, and solidified. Cells were incubated for 3 weeks, and fresh media containing compounds were administered twice a week. Colonies were identified and calculated under an inverted microscope at $\times 100$ magnification.

11. Endothelial tube formation assay

Endothelial tube formation assay was conducted on the Matrigel (BD Biosciences, San Jose, CA), which was added to a 24-well plate in a volume of 100 μ L/well and allowed to be polymerized at 37°C for 30 minutes. Human umbilical vein endothelial cells (HUVECs) at a density of 2×10^4 were seeded into each well and incubated with 0.5 mL of MEDIUM 199 (Sigma-Aldrich) containing 1 % FBS. After stabilized, cells were incubated in the conditioned media driven by A549 cells for 16 hours. Images were captured with OLYMPUS camera attached to the microscope. The tube formation was quantified by measuring the long axis of the individual cells using Image J.

12. Animals and tumor grafts

Male nude mice (BALB/cAnNCrj-nu/nu) were purchased from Charles River

Japan (Shin-Yokohama, Japan) and housed in a specific pathogen-free room. Mice (11 weeks old) were injected subcutaneously in a flank with 1×10^6 LLC1 cells. After tumor volumes reached over 100 mm^3 , mice were intraperitoneally injected with dimethyl sulfoxide (DMSO) or DAS (1 mg/kg) once a day. Tumor volumes were measured with a caliper and calculated using the equation " $\frac{a \times b^2}{2}$ ", where 'a' is the maximal width and 'b' is maximal orthogonal width. All animal procedures were performed in accordance with a protocol approved by the Institutional Animal Care and Use Committee (Approve # SNU-141008-1).

13. Immunohistochemistry

Xenograft tumor tissue specimens were deparaffinized, autoclaved to retrieve antigens, and sequentially incubated with 3 % H_2O_2 , a primary antibody (anti-HIF-1 α , anti-CD31, Thermo Fisher, Rockford, IL; anti-Ki67, Leica Biosystems, Richmond, IL), and a biotinylated secondary antibody (Vector Laboratories, Burlingame, CA). The immune complexes were visualized using Vectastain ABC (Vector Laboratories), and the specimens were counterstained with hematoxylin. Histology and immunostaining were reviewed under a microscope at a $\times 400$ magnification and all slides were reviewed independently by four parts.

14. Statistical analyses

Statistical analyses were performed using the Microsoft Excel (2010) software and values were described as means and standard deviation. The differences between two groups were analyzed by two-sided, unpaired Student t-test, and were considered statistically significant when $P < 0.05$.

RESULTS

Screening of HIF-1 inhibitors

I established the stable Hep3B cell line that expresses luciferase HIF-1 dependently under hypoxia (Fig. 1A) and searched for anti-HIF-1 compounds, as was summarized in Fig. 1B. I first checked chemiluminescence emitting from the cells which had been incubated at 1% oxygen with DMSO or each (finally 5 μ g/mL) of 7,280 compounds in a chemical library for 16 hours. From the first-round screening, I picked up 166 candidates. To rule out the possibility that luciferase expression is nonspecifically impaired due to toxicity, I checked whether the candidates at 5 mg/mL were not toxic (less than 20 % cell death) using MTT analysis. From the second-round screening, 27 compound were selected as HIF-1 inhibitors. To further search for potent inhibitors, I reevaluated the anti-HIF-1 effects of 27 candidates at a concentration of 1 mg/mL and picked up three candidates. Finally, I selected the most potent compound “BHS-A503” that inhibited HIF-1 at < 100 ng/mL. The compound was an unrefined extract from *Bacillus licheniformis*, which is parasitic on red alga *Gelidium pacificum*.

Purification and identification of a HIF-1inhibitor from the BHS-A503 fraction

To purify a single compound from the bacterial extract, I performed silica-gel flash chromatography and separated the extract into 40 fractions. Of them, I picked up the fraction 26 because it inhibited HIF-1 activity by > 85% (Fig. 1C) and was also not significantly toxic (Fig. 1D). The effects of the fractions against HIF-1 were rechecked by evaluating the HIF-1 α protein level using Western blotting. As was expected, HIF-1 α protein was not detected under normoxia but was notably induced under hypoxia. I also found that the fraction 26 at 100 ng/mL attenuated the hypoxic induction of HIF-1 α almost completely (Fig. 1E). Since the fraction 26 was not a single compound, I further separated the fraction 26 into 13 fractions using LH-20 column chromatography. Consequently, the 7th fraction of the fraction 26, which is designated #26-7, was identified not only to inhibit HIF-1 activity even at 10 ng/mL but also to be less toxic (Fig. 2A and 2B). A Western blotting showed that #26-7 suppressed HIF-1 α expression very effectively (Fig. 2C).

Anti-HIF effects of #26-7 in different cancer cell lines

We next examined whether #26-7 has anti-HIF activity in cancer cells other

than Hep3B. #26-7 suppressed HIF-1 α expression in a dose-dependent manner, whereas it did not substantially affect ARNT expression (Fig. 3A). Likewise, #26-7 also effectively suppressed HIF-1 α expression in HCT116 and MKN45 cells (Fig. 3B). Apparently, the efficacy of #26-7 on HIF-1 α inhibition appears to be similar among three cell lines.

In the EPO enhancer reporter assay, #26-7 was found to inhibit the HIF-1 driven transcription under hypoxia (Fig. 3C). To examine whether #26-7 regulates the cellular response to hypoxia, I analyzed mRNAs of the HIF target genes in A549 cells. The hypoxic induction of LOX, CA9, CyclinD1, and CITED2 mRNAs was substantially attenuated by #26-7 (Fig. 3D). Using [α]D, LR-EI-MS, and NMR (^1H , ^{13}C , DEPT) analyses, #26-7 was identified as diacetoxyscirpenol [Cole et al., 1981]. Its structure is illustrated in Fig. 4A. First, I compared the toxicity of the purchased diacetoxyscirpenol in A549 and two normal cell-lines. Even at the higher concentrations of diacetoxyscirpenol, no significant differences in toxicity were observed among the cell-lines (Fig. 4B). Given that this reagent also inhibited the expression and activity of HIF-1/2 α (Fig. 4C and 4D), diacetoxyscirpenol was considered an HIF-inhibiting ingredient included in #26-7. Compared with a known HIF-1 inhibitor 17AAG, diacetoxyscirpenol was shown to be more potent in HIF-1 inhibition (Fig. 4E).

Diacetoxyscirpenol deregulates HIF-1 signaling in two different steps

To understand how diacetoxyscirpenol downregulates HIF-1 α , I first checked the stability of HIF-1 α protein. HIF-1 α degradation was induced by reoxygenation after the protein was accumulated under hypoxia (Fig. 5A) or by cycloheximide treatment after it was stabilized using a HIF prolyl hydroxylase inhibitor, dimethyloxaloylglycine (DMOG) (Fig. 5B). However, the rate of HIF-1 α degradation was not facilitated in the presence of diacetoxyscirpenol, suggesting no effect of diacetoxyscirpenol on HIF-1 α stability. Next, I analyzed de novo synthesis of HIF-1 α using MG132, which can induce the accumulation of newly synthesized HIF-1 α protein by blocking protein degradation. Strikingly, the synthesis of HIF-1 α protein was abolished by diacetoxyscirpenol (Fig. 5C). To further examine the mechanism underlying the blockade of HIF-1 α synthesis, I measured the mRNA levels of HIF-1 α , but diacetoxyscirpenol did not inhibit the transcription of HIF-1 α (Fig. 5D), which encouraged us to analyze the translation of HIF-1 α . The translation of HIF-1 α mRNA is initiated through two pathways, 5' cap-dependent translation and internal ribosome entry site (IRES)-dependent translation, both of which are determined by the 5'-UTR of HIF-1 α mRNA [Lang et al., 2002]. I analyzed the activities of two luciferase reporters reflecting each pathway of HIF-1 α translation. Consequently, diacetoxyscirpenol inhibited both the pathways of HIF-1 α translation, whereas it

did not affect the activity of TK promoter without the 5'-UTR (Fig. 5E). Diacetoxyscirpenol seems to inhibit de novo synthesis of HIF-1 α protein by blocking the 5'-UTR-mediated translation of HIF-1 α mRNA. To rule out other actions of diacetoxyscirpenol on oxygen-dependent HIF-1 regulation, I examined if it affects the HIF-1 α hydroxylation by PHD enzymes or the HIF-1 α ubiquitination/degradation by the pVHL-E3 complex. However, diacetoxyscirpenol suppressed HIF-1 α expression even in the presence of DMOG and also in VHL-null RCC4 cells (Fig. 6A and 6B). These findings further support our notion that diacetoxyscirpenol does not affect the degradation process of HIF-1 α . To get its functionality as a transcription factor, HIF-1 α should dimerize with ARNT in the nucleus. Interestingly, diacetoxyscirpenol interfered with the dimerization of HIF-1 α and ARNT (Fig. 6C), which may be attributed to impaired nuclear translocation of HIF-1 α . The level of nuclear HIF-1 α was cross-checked using immunoblotting (Fig. 6D) and immunofluorescence (Fig. 6E). Collectively, diacetoxyscirpenol seem to attenuate HIF-1 signaling in two different ways.

Diacetoxyscirpenol inhibits anchorage-independent cancer growth and angiogenesis

Anchorage-independent cell growth is a representative property of cancer cells with tumorigenic potential, and it has been reported to occur depending on HIF-1 α [Lee et al., 2013; Mills et al., 2009]. Therefore, I examined if diacetoxyscirpenol can negate the tumorigenic potential of A549 cells using a colony formation assay. At a glance, colonies of A549 were shown at a lesser density in the presence of diacetoxyscirpenol (Fig. 7A). Both the number and size of colonies were significantly reduced by diacetoxyscirpenol in a dose-dependent manner (Fig. 7B). I next investigated if diacetoxyscirpenol attenuates the angiogenic potential of cancer cells because HIF-1 α plays a key role in tumor angiogenesis [Pugh et al., 2003; Chang et al., 2006]. Diacetoxyscirpenol substantially inhibited VEGF secretion from A549 cells in either normoxic or hypoxic conditions (Fig. 8A). As expected, diacetoxyscirpenol attenuated the hypoxic induction of VEGF mRNA (Fig. 8B). To check the angiogenic potential of A549 cells, tube formation of HUVECs was analyzed in the presence of A549-driven conditioned media. Endothelial tube formation was augmented by the conditioned media from A549 cells exposed to hypoxia. However, in the conditioned media from A549 cells treated with diacetoxyscirpenol, the tube formation was significantly reduced (Fig. 8C). To

test the possibility that diacetoxyscirpenol per se inhibits the endothelial tube formation, the compound was separately administered into the endothelial culture plates containing conditioned media from hypoxic A549 cells. Given that the tube formation was attenuated by diacetoxyscirpenol (Fig. 8D), diacetoxyscirpenol is also likely to have a direct effect against vascular formation. These results suggest that diacetoxyscirpenol inhibits tumorigenic and angiogenic potentials of cancer cells under hypoxia.

Diacetoxyscirpenol inhibits the growth of lung carcinoma grafts in mice

To in vivo examine the anticancer activity of diacetoxyscirpenol, this agent was administered into mice bearing Lewis lung carcinoma. When tumor volume reached about 100 mm³, mice were intraperitoneally injected with DMSO (vehicle) or diacetoxyscirpenol. No substantial changes in gross appearance and body weight suggest that diacetoxyscirpenol at the used dose is not highly toxic (Fig. 9A). Diacetoxyscirpenol retarded tumor growth (Fig. 9A), and excised tumors were noticeably smaller in diacetoxyscirpenol-treated mice than in vehicle-treated mice (Fig. 9B). The HIF-1 α suppression by diacetoxyscirpenol was confirmed by immunoblotting HIF-1 α in tumor homogenates (Fig. 9C). HIF-1 α and Ki-67 (a marker for cell proliferation) were less expressed, and

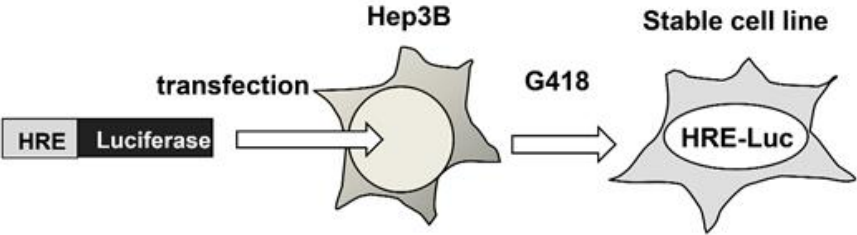
vessels were poorly developed in tumor specimens from diacetoxyscirpenol-treated mice (Fig. 9D and 9E). These results suggest that diacetoxyscirpenol attenuates cancer cell proliferation by impairing tumor adaptation to hypoxic microenvironment. However, given that the numbers of TUNEL-stained apoptotic cells were not different between two groups, this agent may not kill cancer cells directly.

Figure 1. The 1st round screening of anti-HIF compounds in BHS-A503.

(A) Scheme for making Hep3B stable cell lines harboring the EPO enhancer/luciferase reporter. (B) Scheme for searching for compounds having anti-HIF activity. (C) Effects of BHS-A503 fractions against HIF-driven transcription. EPO-Hep3B cells were treated with one of 40 fractions (100 ng/mL) from BHS-A503 for 4 hours, and then incubated at 1% oxygen for 16 hours. Luciferase activities in cell lysates were measured using a luminometer. (D) Effects of BHS-A503 fractions on cell viability. Hep3B cells were treated with 100 ng/mL of each fraction for 24 hours, and cell viabilities were evaluated by MTT assay. (E) Effects of BHS-A503 fractions on HIF-1 α expression. Hep3B cells were treated with 100 ng/mL of each fraction under hypoxia for 12 hours. HIF-1 α and β -tubulin levels were analyzed by immunoblotting. All results were obtained from 4 independent experiments and each bar represents the mean + standard deviation.

Figure 1.

A



B

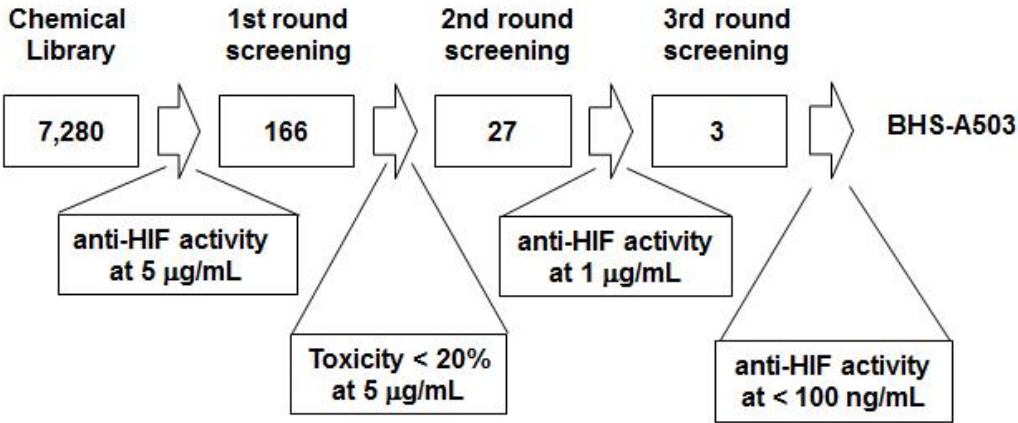
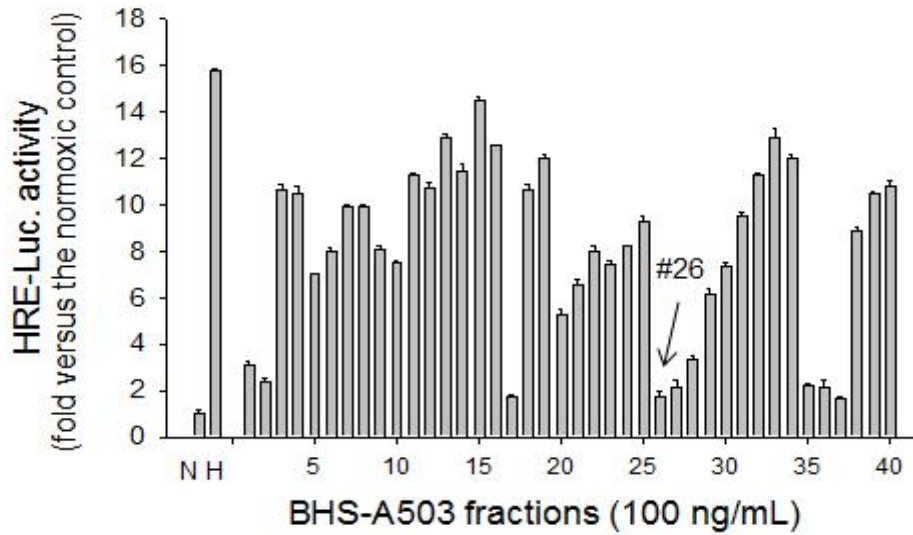
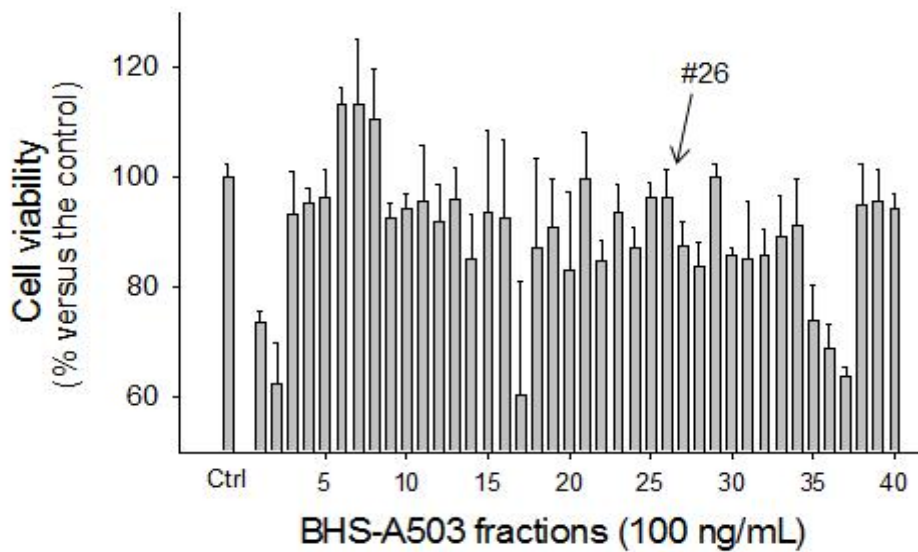


Figure 1. Continued.

C



D



E

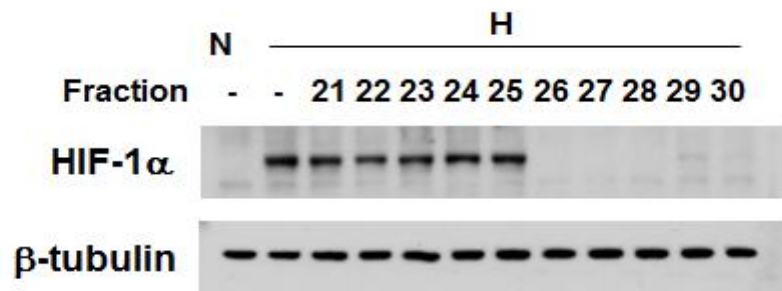
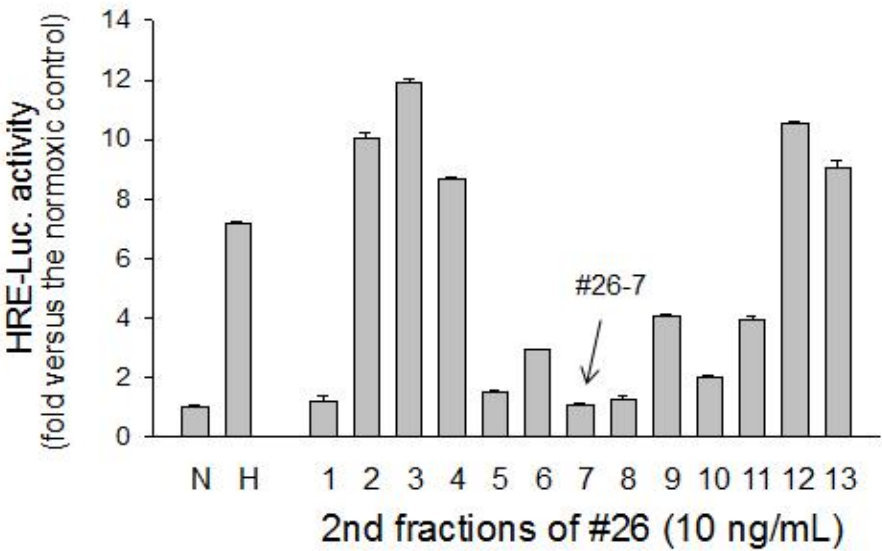


Figure 2. The 2nd round screening of anti-HIF compounds in BHS-A503 #26 fractions.

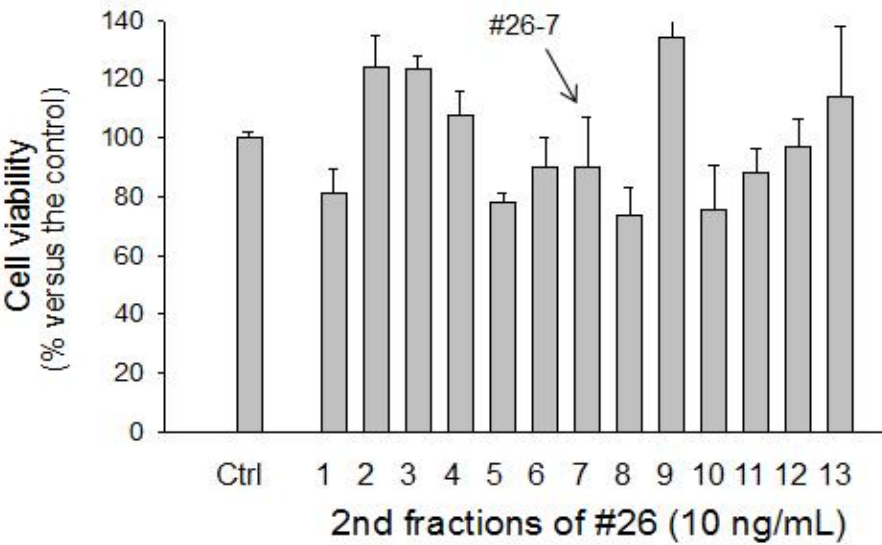
(A) Effects of #26 fractions against HIF-driven transcription. The EPO enhancer/luciferase-harboring Hep3B cells were treated with one of 13 fractions (10 ng/mL) from BHS-A503 #26 for 4 hours, and then incubated under hypoxia for 16 hours. Luciferase activities in cell lysates were measured using a luminometer. (B) Effects of BHS-A503 #26 fractions on cell viability. Hep3B cells were treated with 10 ng/mL of each fraction for 24 hours, and cell viabilities were evaluated by MTT assay. (C) Effects of BHS-A503 #26 fractions on HIF-1 α expression. Hep3B cells were treated with 10 ng/mL of each fraction under hypoxia for 12 hours. HIF-1 α and β -tubulin levels were analyzed by immunoblotting. All results were obtained from 4 independent experiments and each bar represents the mean and standard deviation.

Figure 2.

A



B



C

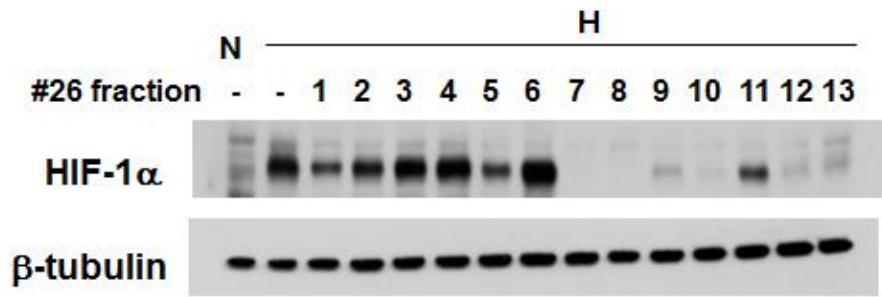


Figure 3. Anti-HIF activity of compound #26-7.

(A and B) A549, HCT116, or MKN45 cells were pre-treated with #26-7 for 4 hours, and further incubated under hypoxia for 8 hours. Protein levels were analyzed by immunoblotting (top). Each bar represents the mean and standard deviation ($n = 3$), and *and # denote $P < 0.05$ versus the hypoxic control (bottom). (C) A549 cells, which had been co-transfected with the EPO enhancer-luciferase and the CMV promoter-galactosidase plasmids (1 mg of each), were treated with #26-7 for 4 hours, and then incubated under hypoxia for 16 hours. Luciferase activities were normalized to galactosidase activities, which were presented as relative values to the normoxic control level. Each bar represents the mean + standard deviation ($n = 4$) and *denotes $P < 0.05$ versus the hypoxic control (the 2nd bar). (D) Quantitative RT-PCR was performed to check the expression of HIF-targeted genes in A549 cells which were treated with #26-7 under normoxia (N) or hypoxia (H) for 16 hours. Each assay was done in triplicate and the result was divided by β -actin level in the corresponding sample. Each bar represents the mean and standard deviation ($n = 4$) and *denotes $P < 0.05$ versus the hypoxic control.

Figure 3.

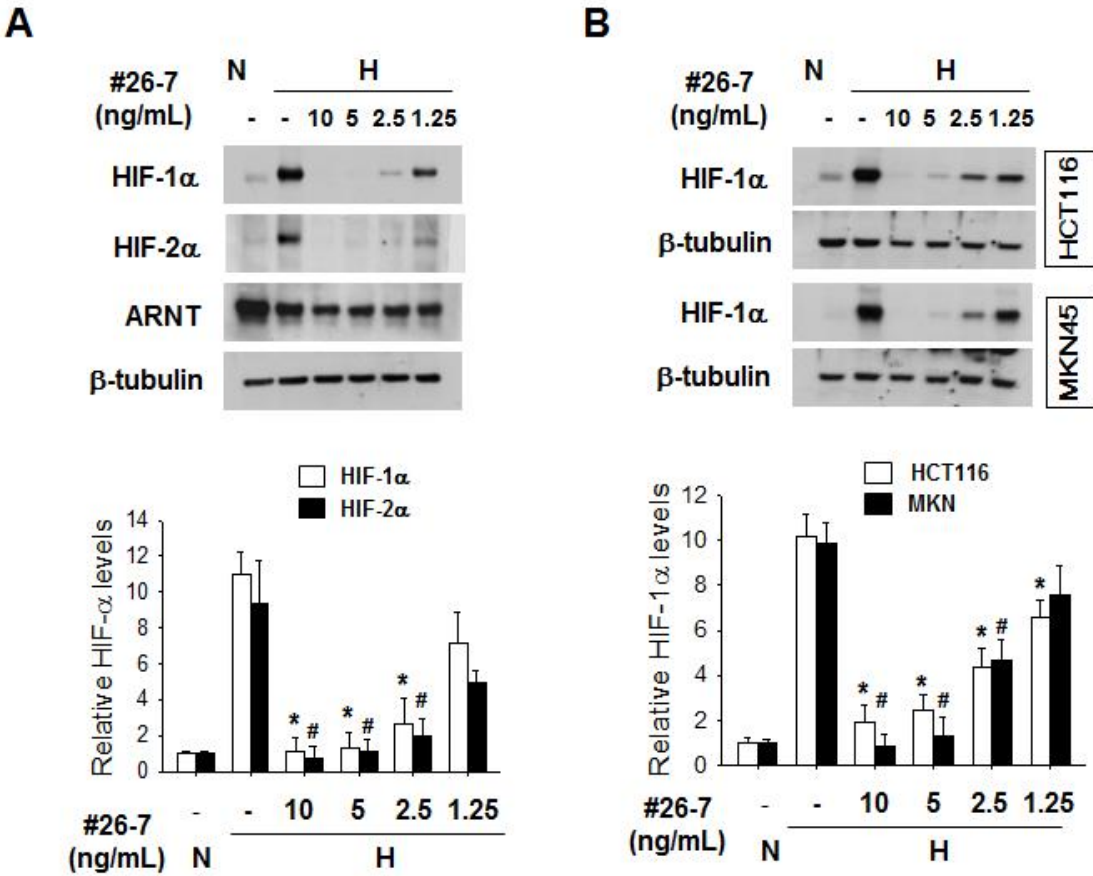
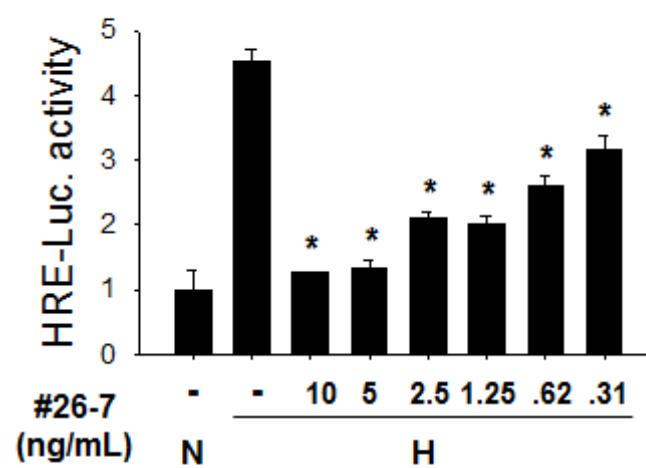


Figure 3. Continued.

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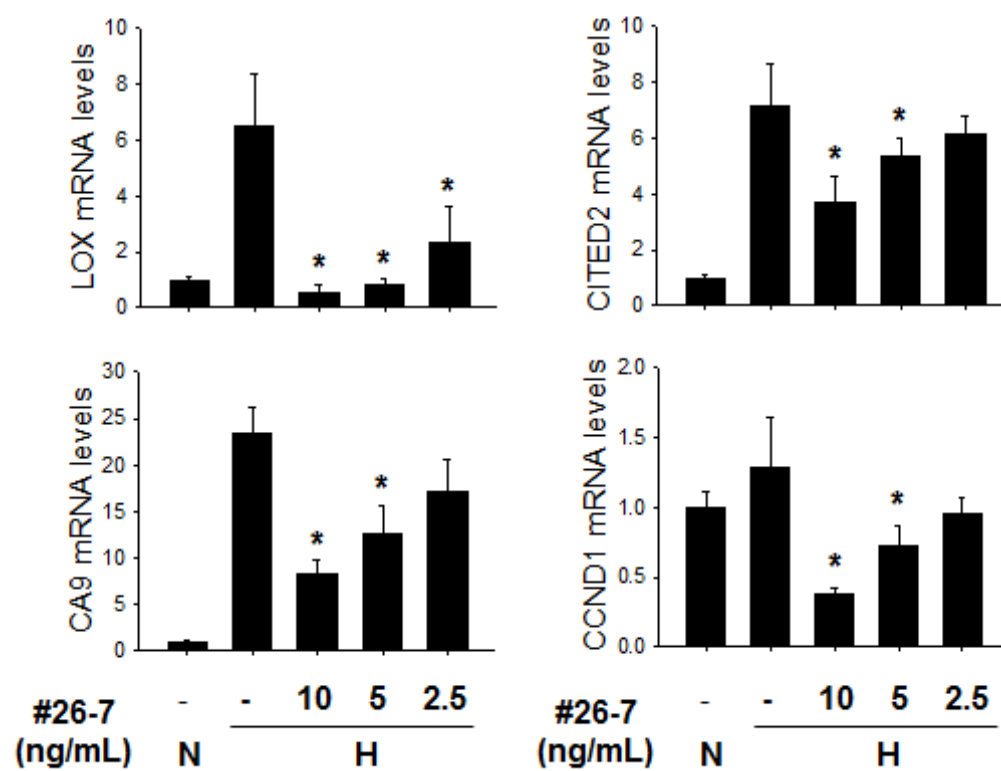


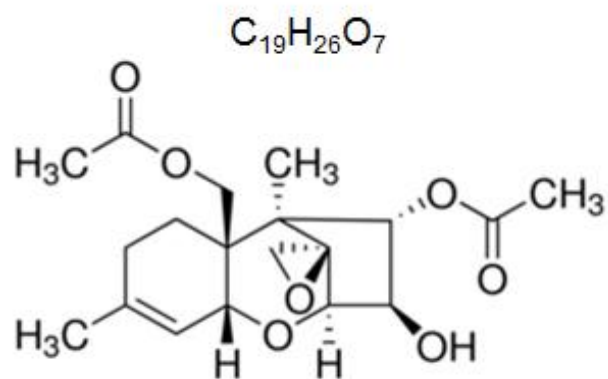
Figure 4. Identification of BHS-A503 #26-7.

(A) Compound #26-7 was identified as diacetoxyscirpenol (DAS) in $[\alpha]$ D, LR-EI-MS, and NMR (^1H , ^{13}C , DEPT) analyses. (B) Comparison of toxicities of DAS among cancer cell and normal cells. A549, CCD-18Lu, and HEK293 were treated with the indicated doses of DAS for 24 hours. Cell viability was evaluated by MTT assay, and each bar represents the mean + standard deviation from 3 independent experiments. (C) A549 cells were treated with 1.25 - 10 ng/mL of DAS, which was purchased from a company, for 4 hours and subjected to 8 hour-hypoxia. Protein levels were analyzed by immunoblotting. Each bar represents the mean and standard deviation ($n = 3$), and * and # denote $P < 0.05$ versus the hypoxic control. (D) A549 cells, which had been co-transfected with EPO enhancer-luciferase and galactosidase plasmids (1 mg of each), were treated with DAS for 4 hours and incubated under hypoxia for 16 hours. Luciferase activities were normalized to galactosidase activities and are presented as relative values to the normoxic control. Each bar represents the mean and standard deviation ($n = 4$) and *denotes $p < 0.05$ versus the hypoxic control. (E) A549 cells harboring the EPO-Luc plasmid were treated with DAS or 17AAG at the indicated concentrations for 4 hours, and incubated under hypoxia for 16 hours. Luciferase activities in drug-treated groups were divided

by those in drug-free group. Each bar represents the mean and standard deviation ($n = 4$), and the half maximal inhibitory concentration (IC_{50}) was calculated using the ED 50 plus program.

Figure 4.

A



B

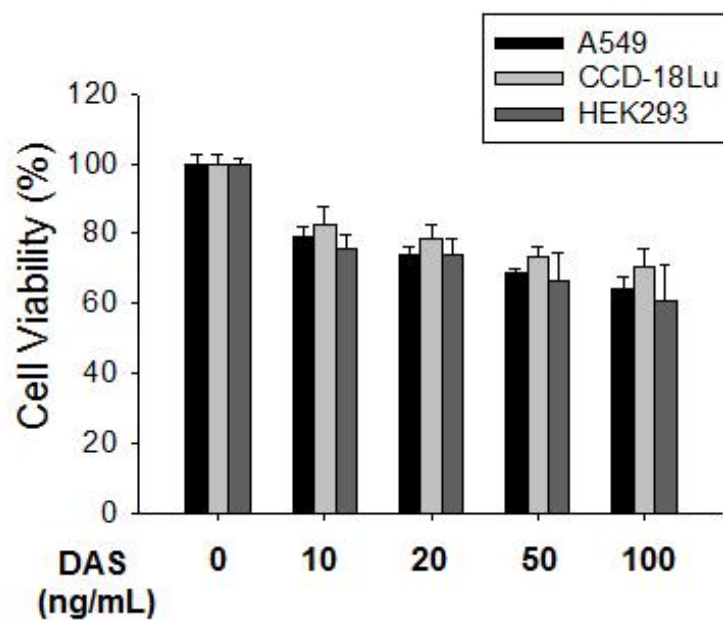
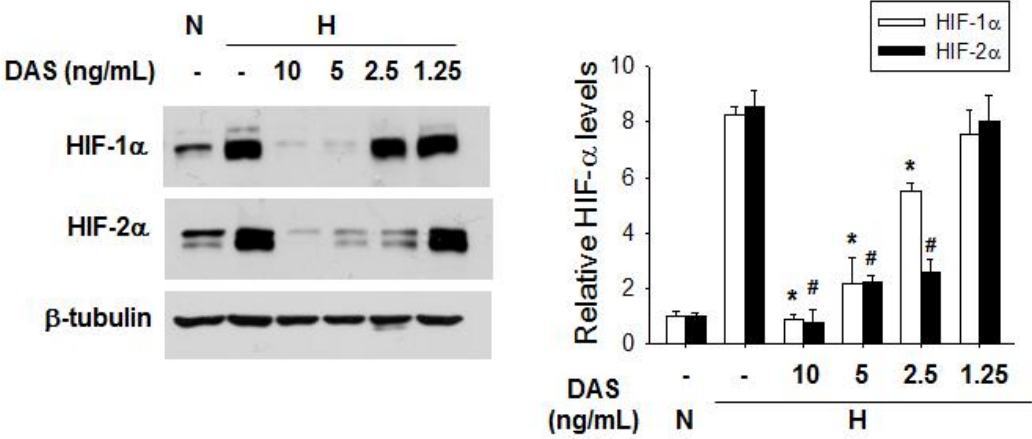
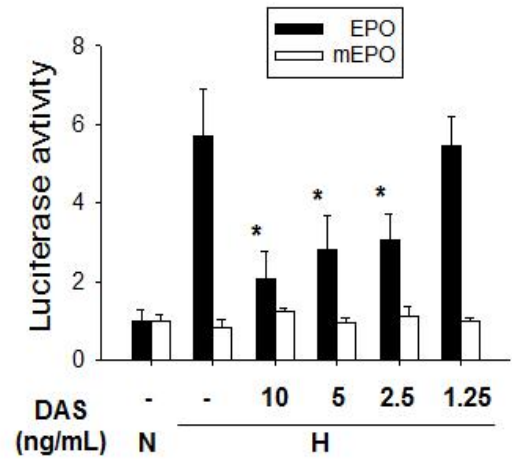


Figure 4. Continued.

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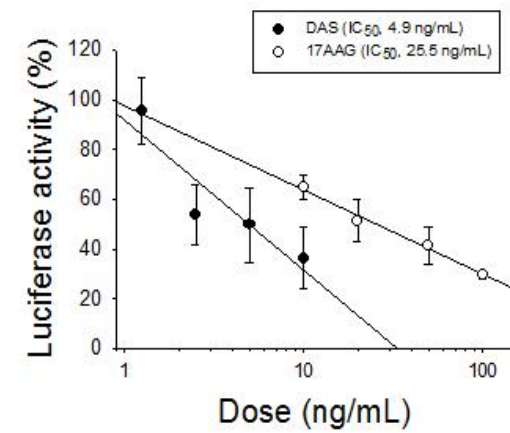


Figure 5. Diacetoxyscirpenol downregulates HIF-1 α in the translational level.

(A) Effect of DAS on oxygen-dependent degradation of HIF-1 α . A549 cells were incubated with DMSO or DAS under hypoxia for 12 hours. Following re-oxygenation for the indicated periods (0 to 10 minutes), cells were lysed and HIF-1 α levels were analyzed by immunoblotting. (B) After treated with 500 μ M DMOG for 8 hours, A549 cells were pre-incubated with DMSO or DAS (10 ng/mL) for 1 hour, and then treated with 10 mg/mL of CHX for the indicated times. HIF-1 α levels were determined by immunoblotting. (C) Effect of DAS on de novo synthesis of HIF-1 α protein. After treated with DMSO or DAS 4 hours, A549 cells were additionally treated with 10 μ M MG132 to induce the accumulation of newly synthesized HIF-1 α . HIF-1 α levels were determined at the indicated times (0 to 8 hours) by immunoblotting (top). Each bar represents the mean and standard deviation ($n = 3$), and *denotes $P < 0.05$ versus the DMSO control at the corresponding time (bottom). (D) Effect of DAS on the transcription of the HIF-1 α gene. Quantitative RT-PCR was performed to check the HIF-1 α mRNA level in A549 cells which were treated with DAS for 16 hours. Each assay was done in triplicate and the result was divided by β -actin level in the corresponding sample. Each bar represents the mean and standard

deviation (n = 4). (E) Effect of DAS on the 5'UTR-driven translation of HIF-1 α . Cap-dependent and IRES-dependent translational rates of HIF-1 α were analyzed using TK promoter-5' UTR-luciferase and CMV promoter-GFP-5'UTR-luciferase reporters, respectively. The TK promoter-luciferase vector was used as a background control. A549 cells, which had been co-transfected with one of these reporters and galactosidase plasmids, were subjected to 16 hour-hypoxia in the absence or presence of DAS. Luciferase activities were normalized to galactosidase activities and are presented as relative values to the normoxic control. Each bar represents the mean and standard deviation (n = 4) and *denotes $p < 0.01$ versus the hypoxic control.

Figure 5.

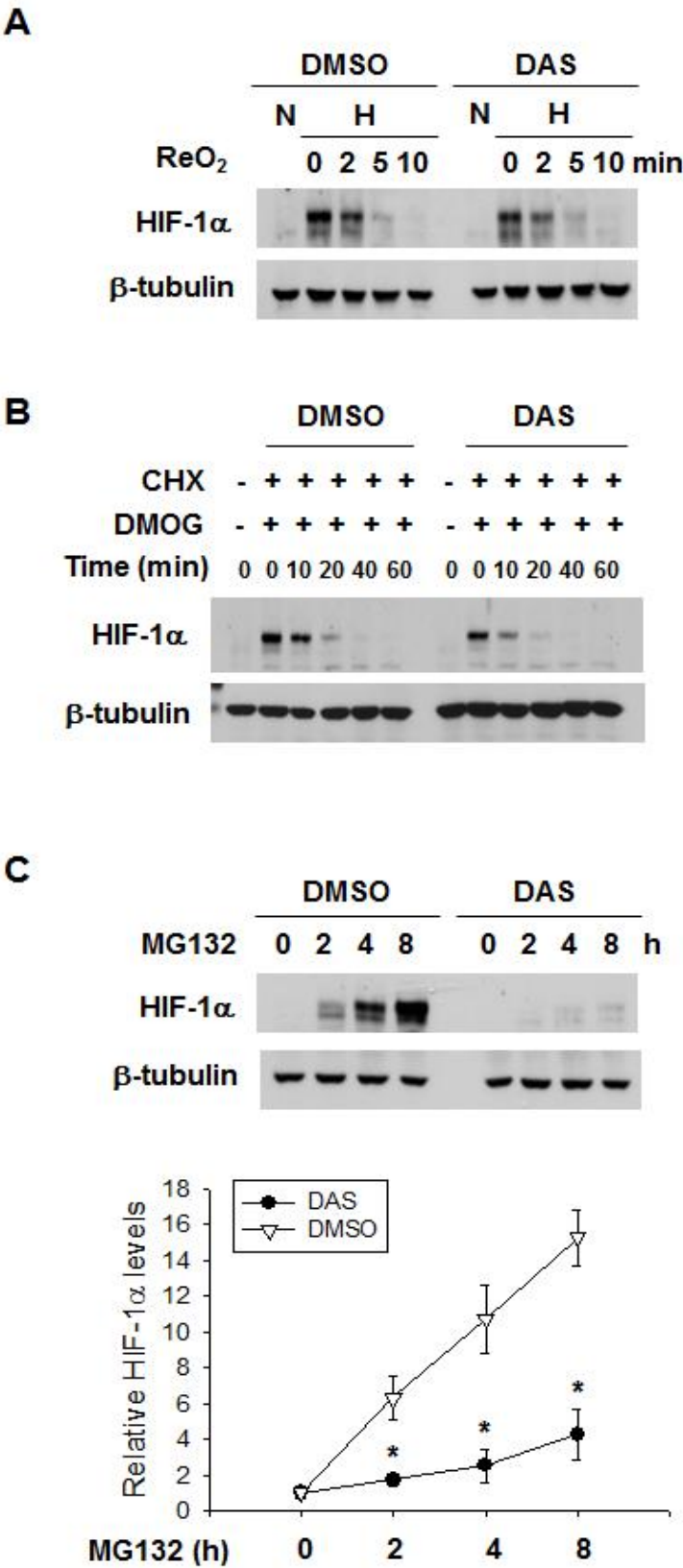


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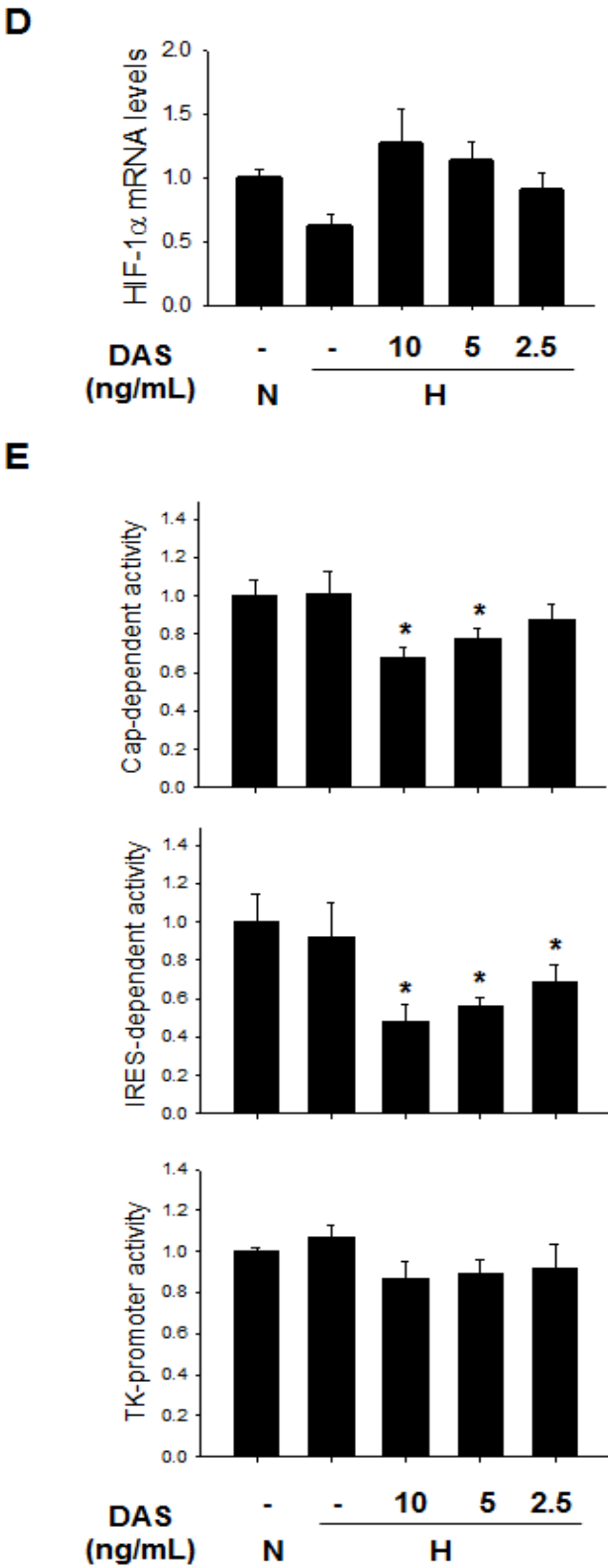


Figure 6. Diacetoxyscirpenol blocks the formation of the HIF-1 complex.

(A) After treated with DAS for 4 hours, A549 cells were incubated with 0.5 mM DMOG to inhibit the hydroxylation of HIF-1 α , and subjected to immunoblotting. (B) VHL-null RCC4 cells were treated with DAS for 12 hours, and subjected to immunoblotting. (C) A549 cells were pretreated with DAS (5 ng/mL) for 4 hours, and further incubated under hypoxia for 8 hours. HIF-1 α or ARNT were precipitated with the indicated antibodies, and then, the precipitates was subjected to immunoblotting. The specificity of immunoprecipitation was verified using non-immunized serum (IgG). (D) The lysates of A549 cells, which had been treated as indicated, were separated into the cytosolic and nuclear fractions, which were subjected to immunoblotting. Lamin B and β -tubulin were analyzed to verify the specificity of each fraction. (E) A549 cells were treated DMSO or DAS (5 ng/mL) under normoxic or hypoxic conditions for 16 hours. The cells were co-stained with anti-HIF-1 α antibody (green) and DAPI (blue). The fluorescence images were captured at $\times 400$ magnification under a fluorescent microscope.

Figure 6.

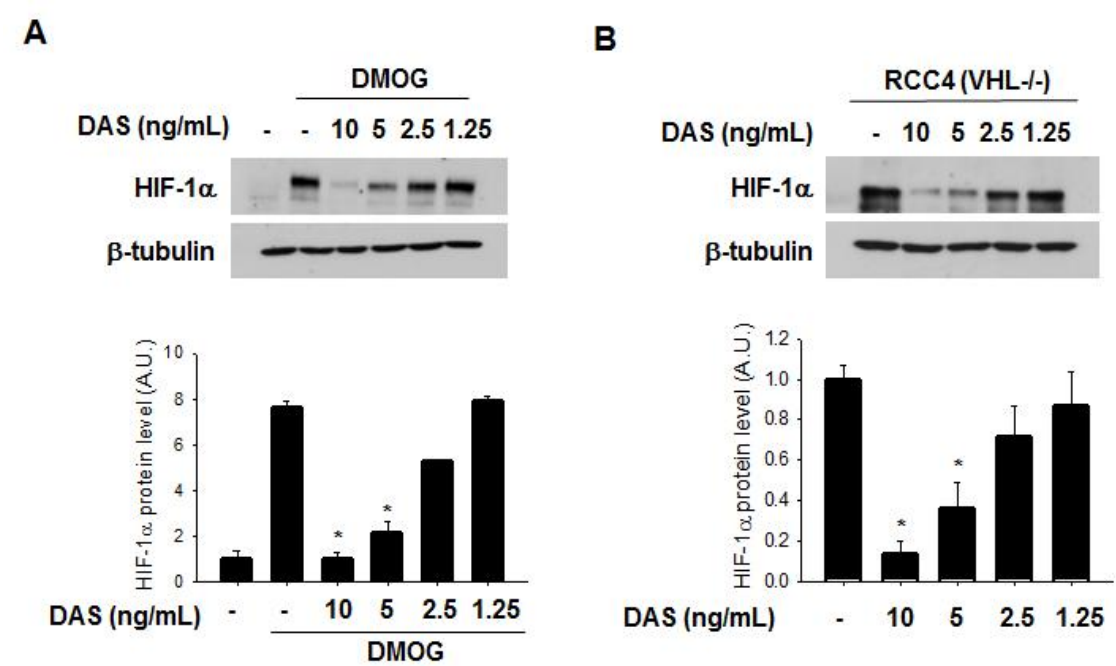


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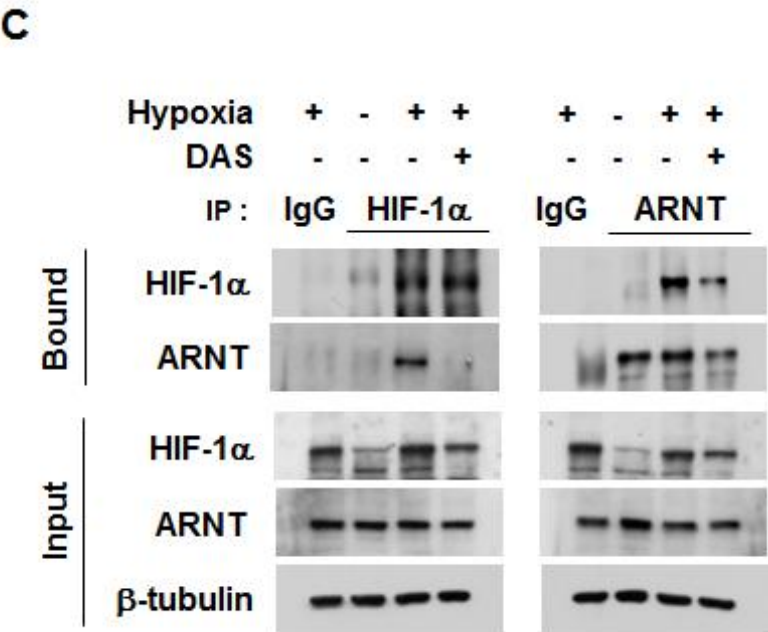
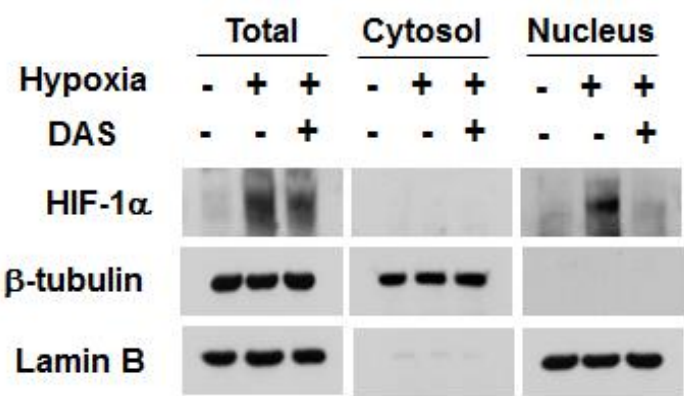


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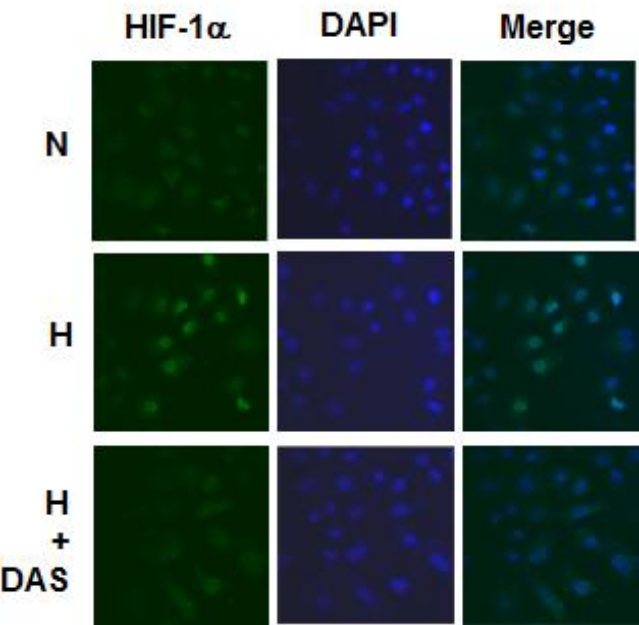


Figure 7. Effects of diacetoxyscirpenol on the colonial growth of A549 cancer cells.

(A) After treated with DAS twice a week for 3 weeks, A549 cells in the top agar were stained with 0.05 % crystal violet. (B) Colony size was measured using Image J and colonies whose sizes were over 0.2 mm^2 were counted. Each bar represents the mean + standard deviation ($n = 4$) and *denotes $P < 0.05$ versus the DMSO control.

Figure 7.

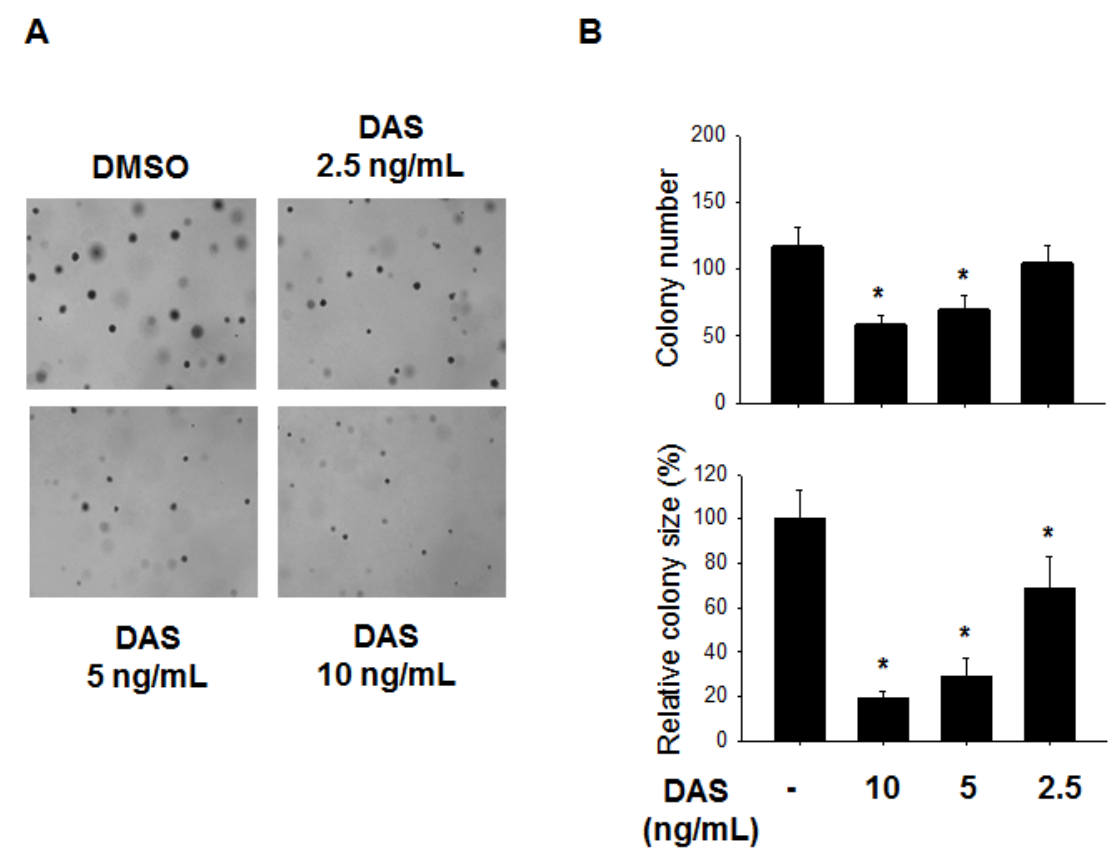


Figure 8. Anti-angiogenic effect of diacetoxyscirpenol.

(A) The conditioned media were collected from A549 cells which had been incubated with DAS under normoxic or hypoxic conditions. The concentrations (means + standard deviation, $n = 4$) of VEGF in the conditioned media were measured using an ELISA kit. * and # denote $P < 0.05$ versus the normoxic control and the hypoxic control, respectively. (B) After A549 cells were treated with DAS for 24 hours under hypoxia, the VEGF mRNA levels were analyzed by quantitative RT-PCR. Each assay was done in triplicate and the VEGF level was divided by β -actin level in the corresponding sample. Each bar presents the mean + standard deviation ($n = 4$) of relative values and * denotes $P < 0.05$ versus the hypoxic control. (C) HUVECs were cultured on Matrigel™-coated plates for 16 hours in the conditioned media from A549 cells which had been treated with DAS for 24 hours under normoxic or hypoxic conditions. The reorganization of HUVECs was pictured under an inverted microscope. Quantitative analyses of tube lengths (mm) and tube area (% of total area) were performed using Image J. (D) After A549 cells were incubated for 24 hours under normoxic or hypoxic conditions, conditioned media were collected and applied to HUVECs in Matrigel™-coated plates. HUVECs were further incubated with DMSO or DAS for 16 hours. HUVECs were photographed and

endothelial tubes were quantified using Image J. Each bar presents the mean + standard deviation (n = 4) and *denotes $P < 0.05$ versus the hypoxic control.

Figure 8.

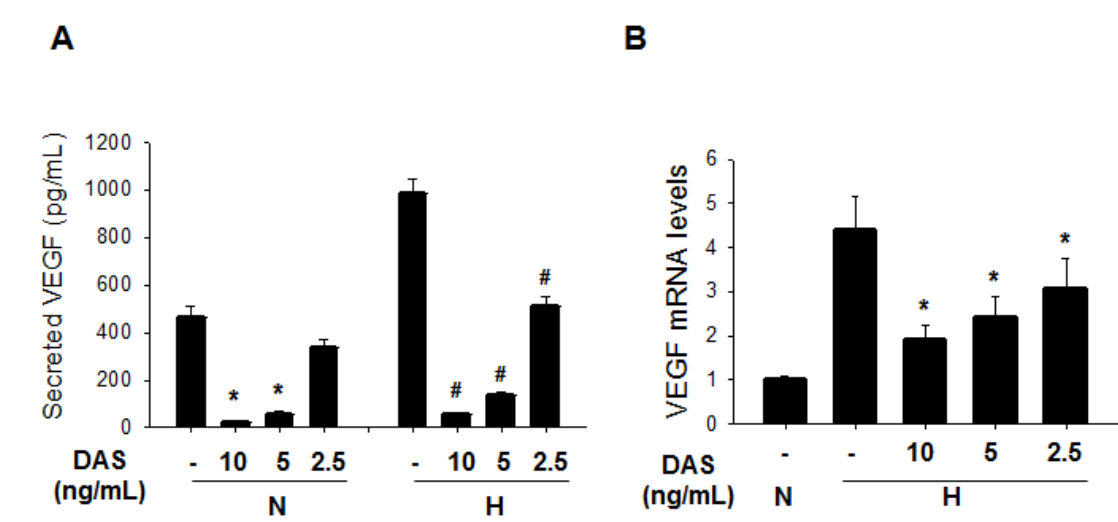


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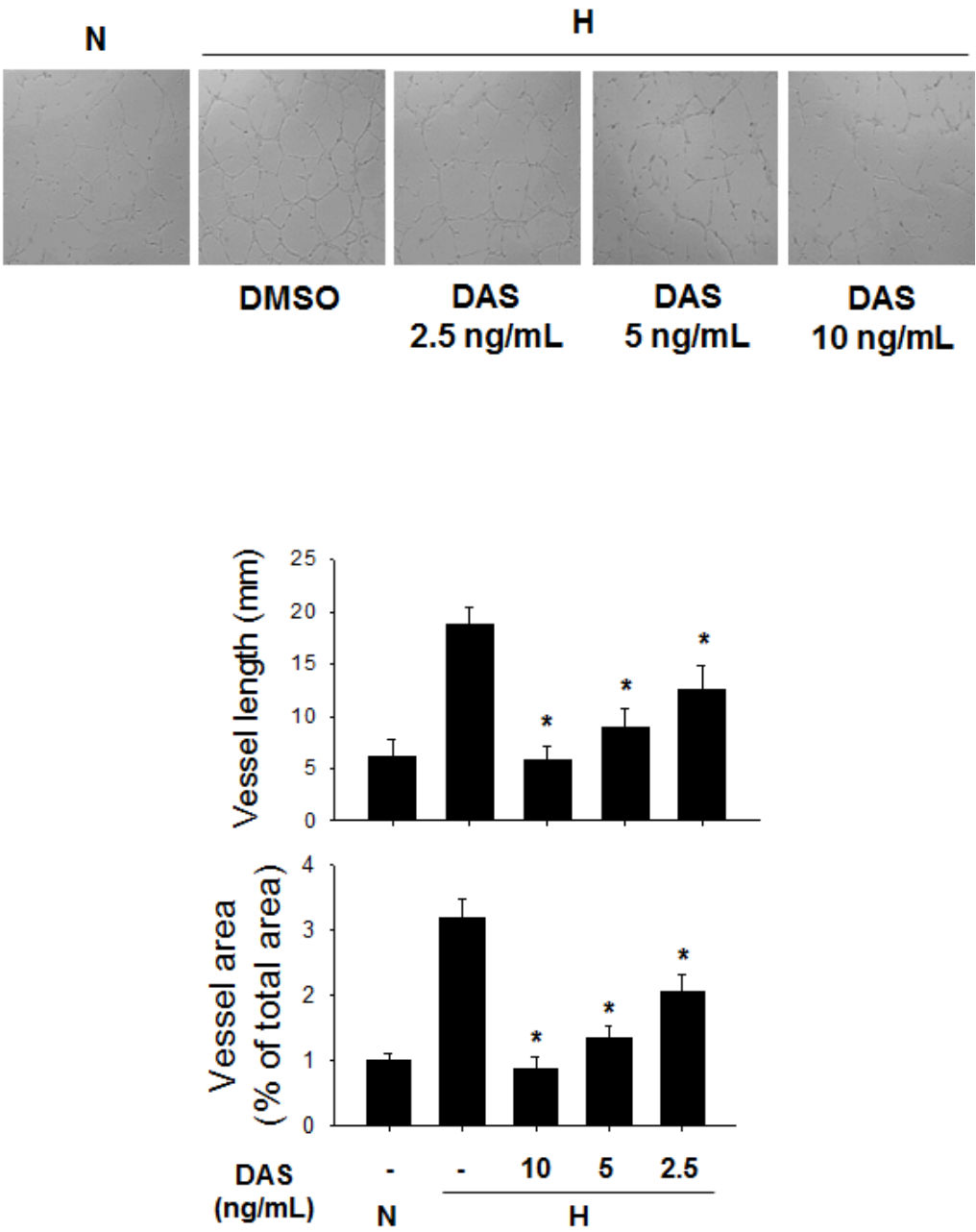


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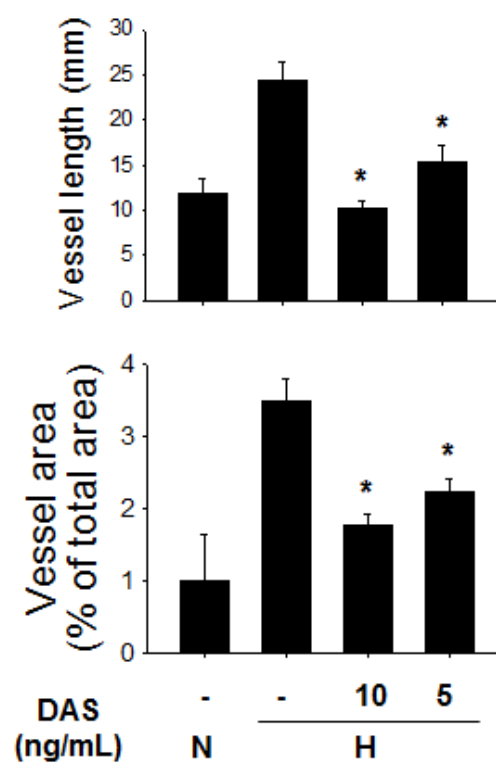
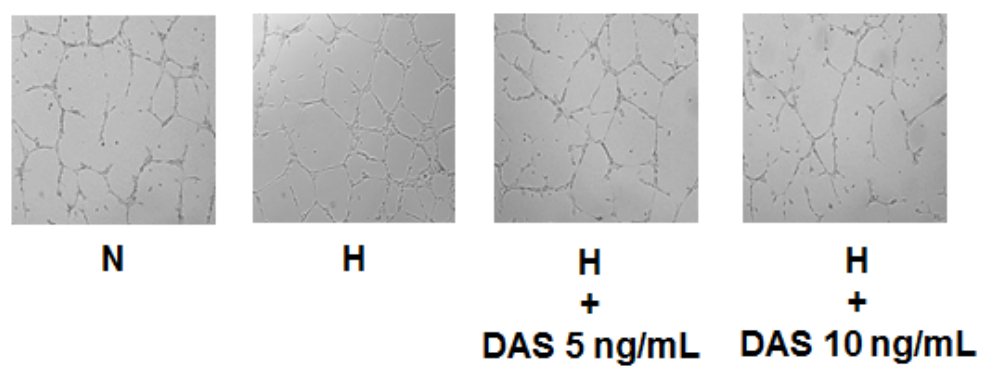
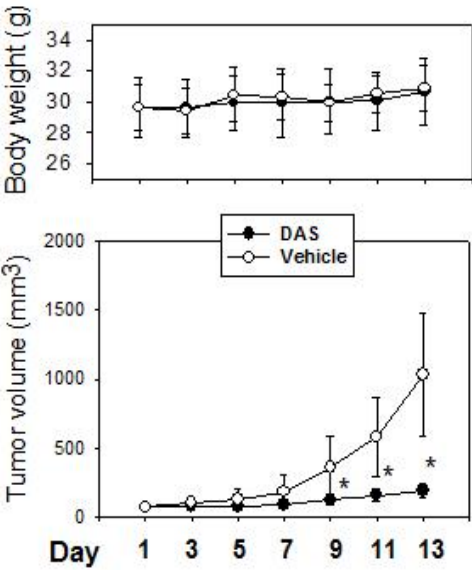


Figure 9. Anticancer effect of diacetoxyscirpenol.

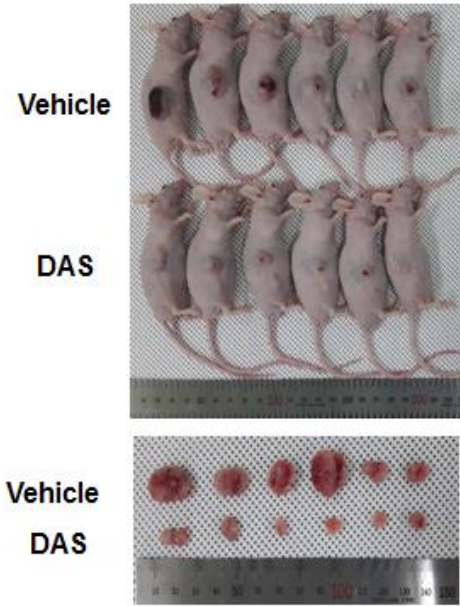
(A) Tumor growth curves. LCC1 cells were grafted on the flanks of mice, and the tumor-bearing mice were injected daily with vehicle (n = 6) or 1 mg/kg of DAS (n = 6) for 2 weeks. Each circle represents the mean and standard deviation of body weight (top) or tumor volume (bottom). *denotes $P < 0.05$ vs. the vehicle group. (B) Gross appearance of tumorbearing mice or excised tumors. (C) In tumor homogenates, HIF-1 α protein was immunoblotted (left), its level was analyzed using ImageJ (right). Each bar represents the mean + standard deviation (n = 5) and *denotes $P < 0.05$ versus the vehicle control. (D) Immunohistochemical analyses. Excised tumors were fixed, paraffin-embedded, and cut into 4 mm sections. The specimens were stained with hematoxylin and eosin (H&E), or immunostained with anti-HIF-1 α , anti-CD31 and anti-Ki-67 antibodies. TUNEL staining was performed to detect cancer cells undergoing apoptosis. (E) Four fields (x400 magnification) per section and four sections of each tumor were reviewed for histologic assessments. HIF-1 α , CD31, Ki-67, TUNEL-positive cells were counted, and the areas of CD31-positive vessel were measured using the Image J. Each bar represents the mean + standard deviation (n = 6) and *denotes $P < 0.05$ versus the vehicle group. Scale bar = 100 μ m.

Figure 9.

A



B



C

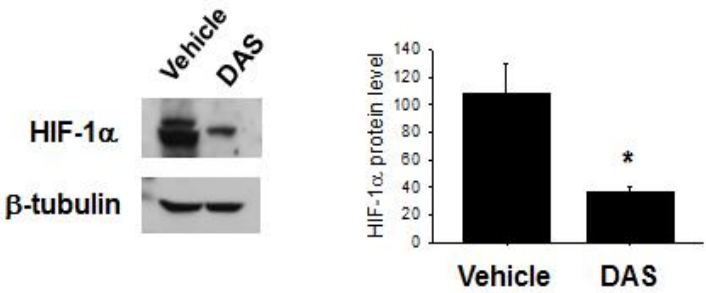
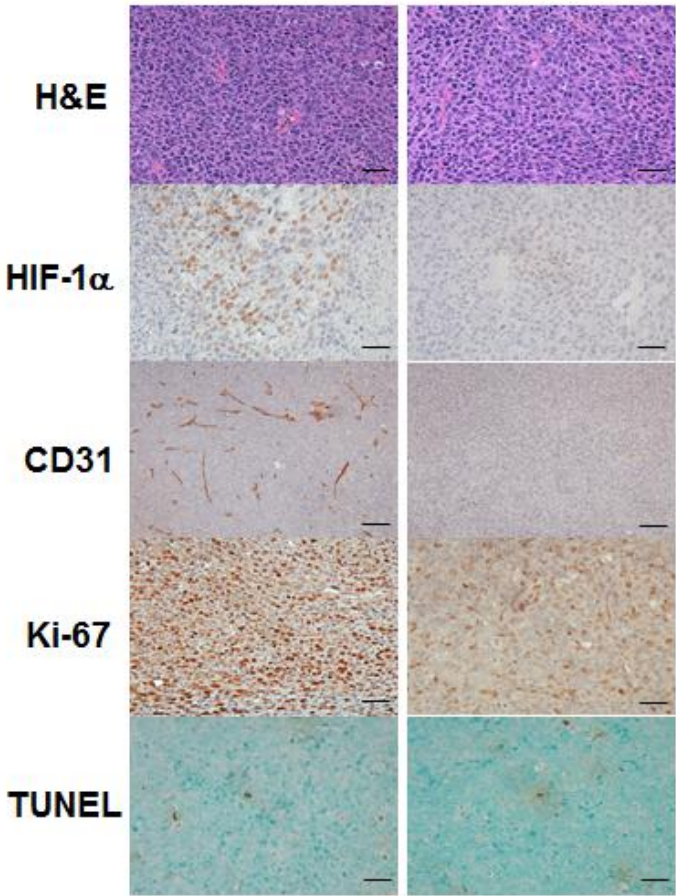


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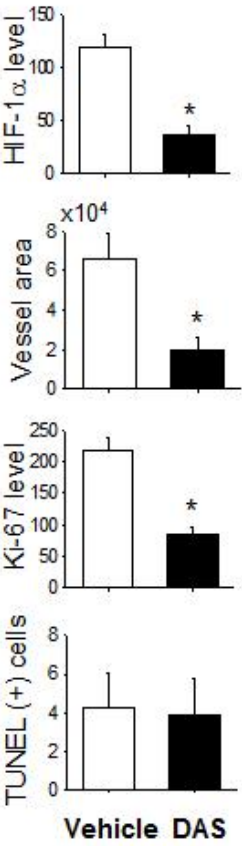
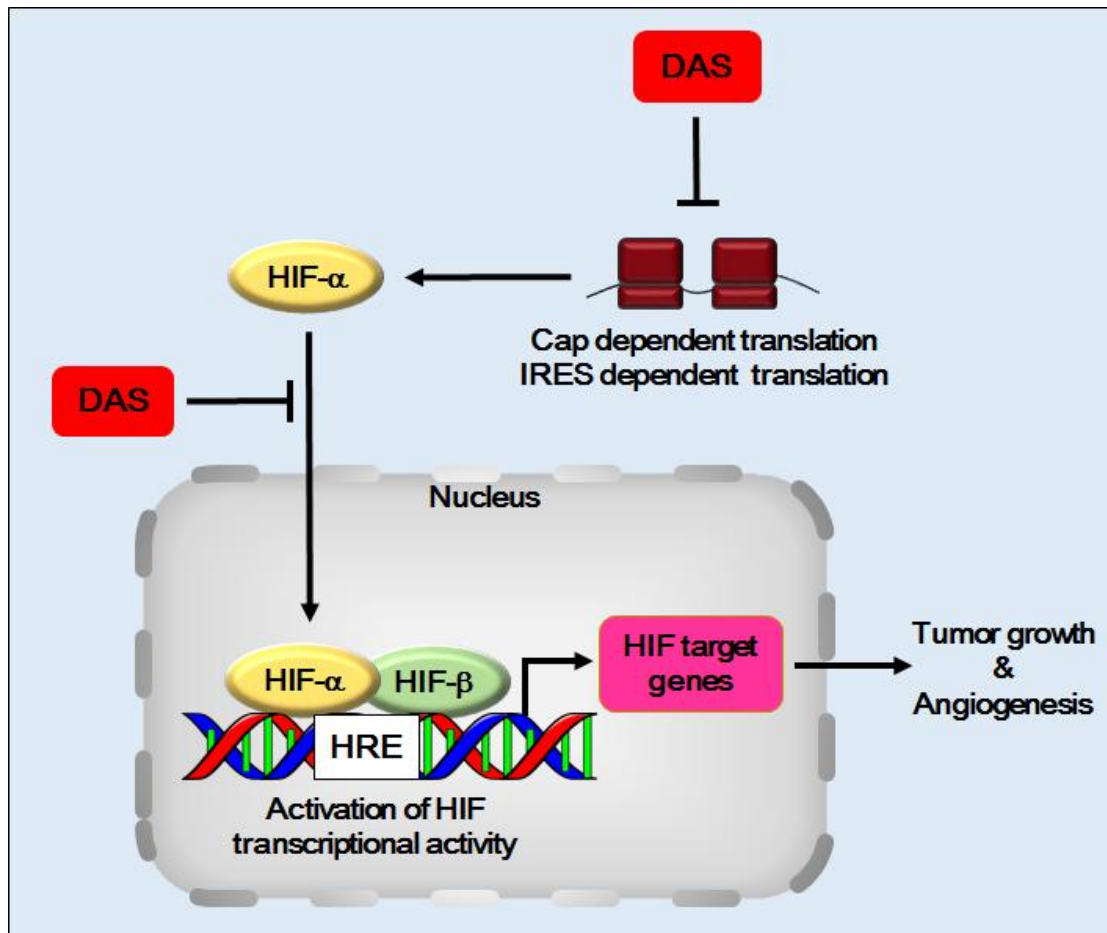


Figure 10. Schematic summary of the role of diacetoxyscirpenol on HIF-1.



DISCUSSION

To find a new anticancer small molecule to target HIF-1, I performed the high-throughput screening using a luciferase reporter system reflecting HIF-driven transcription, and finally selected a marine bacterial extract based on high potency on HIF inhibition and less cytotoxicity. Through two fractionation processes of the extract, diacetoxyscirpenol was purified and identified as an active ingredient having anti-HIF activity. Mechanistically, diacetoxyscirpenol repressed HIF-1 by inhibiting either HIF-1 α translation or its dimerization with ARNT (Fig. 10). Furthermore, diacetoxyscirpenol blunted HIF-mediated induction of hypoxic genes, and by doing so attenuated anchorage-independent colony formation, endothelial tube formation, and tumor growth in mice. Therefore, I propose that diacetoxyscirpenol be a potential anticancer agent targeting HIF-1. To search for compounds inhibiting the HIF-1 signaling pathway, I performed two different methods to evaluate this pathway, namely, the HRE-luciferase reporter and Western blotting for HIF-1 α . However, as shown in Fig. 1C and 1E and in Fig. 2A and 2C, the results from the two analyses were somewhat mismatched with each other.

Actually, the functionality of HIF-1 is determined by diverse factors other than

the amount of HIF-1 α protein. For instance, HIF-1 α is modified (i.e., hydroxylated, phosphorylated, sumoylated, and neddyated), and interacts with many proteins, thereby modulating the HIF-1 activity [Brahimi et al., 2005; Dengler et al., 2014]. The discrepancies between the results from two analyses may be attributed to the post-translation regulations of HIF-1 α . Therefore, I chose the compounds that inhibited HIF-1 at both the analyses. Diacetoxyscirpenol contains the 12,13-epoxytrichothecene group of sesquiterpenes as the core structure. It has been known as one of harmful metabolites produced from plant parasitic fungi belonging to *Fusarium* species [Dawkins et al., 1996]. Accidentally, people or livestock have been subjected to food poisoning by diacetoxyscirpenol which contaminated agricultural crops, such as grains, potatoes, peas, soybeans [Egmond et al., 2007]. However, diacetoxyscirpenol has been rarely reported to contaminate sea plants. Surprisingly, diacetoxyscirpenol was found to be also contained in a marine bacterium parasitizing red alga, suggesting that diacetoxyscirpenol probably cause food poisoning derived by sea foods. For keeping sea foods safe, it may be recommended to check the contamination of diacetoxyscirpenol.

Diacetoxyscirpenol and its analogues were developed as anti-cancer agents because it inhibited cell cycle progression and cell survival [Jun et al., 2007]. Diacetoxyscirpenol was also reported to potentiate the anti-cancer effect of

cisplatin [Hromas et al., 1986]. However, the clinical trial of diacetoxyscirpenol for cancer treatment was ceased at the phase II because it induced intolerable side effects, such as nausea, vomiting, fever, hypertension, and confusion [Bukowski et al., 1982; Yap et al., 1979]. Nonetheless, I argue that diacetoxyscirpenol can be repositioned as an anti-HIF agent. Compared to the concentrations (> 100 nM) of diacetoxyscirpenol used to kill cancer cells directly [Dosik et al., 1978], a lower concentration (< 5 nM) of diacetoxyscirpenol was found in our experiments to inhibit HIF-1. Indeed, cell viability was not substantially decreased by diacetoxyscirpenol at this concentration. If its dose is readjusted to a HIF-inhibiting level, diacetoxyscirpenol is expected to come out as a potential drug that specifically blocks hypoxia-triggered, malignant cancer behaviors, such as angiogenesis, metastasis, and drug resistance.

Diacetoxyscirpenol was shown to block the hypoxia induced production of VEGF, which is a target gene of HIF-1. VEGF does not only drives angiogenesis but also acts as a cell survival factor, which is why VEGF inhibitors have been widely used as anti-cancer drugs. For example, bevacizumab interferes with VEGF binding to the VEGF receptors by sequestering VEGF, and inhibitors of VEGF receptor tyrosine kinase block the signaling pathway of the VEGF receptors [Ferrara et al., 2004; Press et al., 2007]. Diacetoxyscirpenol inhibited the VEGF secretion from cancer cells under

hypoxia, thereby inhibiting the endothelial tube formation. Although it does not directly target endothelial cells, diacetoxyscirpenol may indirectly inhibit tumor-driven angiogenesis at least under hypoxic microenvironment. Some anti-angiogenic agents, mostly in combination with conventional therapeutics, have been shown to have anticancer activity in preclinical and clinical studies [Ma et al., 2008]. However, the pros and cons of anti-angiogenic agents have been debated so far. In some preclinical studies, co-administration of anti-angiogenic agents was shown to aggravate tumor hypoxia and also to impair the delivery of cancer cell-targeting drugs, leading to acquired resistance to chemotherapy [Huang et al., 2015]. By contrast, beneficial actions of anti-angiogenic agents in combination therapies have been also suggested. These agents may improve oxygen and drug delivery within tumors through normalization of irregular tumor vessels [Carmeliet et al., 2011]. Since the anti-angiogenic action of diacetoxyscirpenol may contribute to its anticancer effect, this conflicting point should be considered in developing diacetoxyscirpenol as an adjuvant cancer drug.

In conclusion, I found that diacetoxyscirpenol deregulates tumor adaptation to hypoxia by deregulating HIF-1 α . To our best knowledge, diacetoxyscirpenol is for the first time identified to inhibit HIF-1. More importantly, it is possible that diacetoxyscirpenol is used as a HIF-1 inhibitor at a lesser dose than its

cytotoxic dose. If a low dose of diacetoxyscirpenol is limitedly used for the purpose of preventing HIF-mediated tumor progression or drug resistance, it may be worthwhile reevaluating this agent as an adjuvant cancer drug.

REFERENCES

Bardos, J.I and Ashcroft, M. Negative and positive regulation of HIF-1: a complex network. *Biochim Biophys Acta*. 2005; 1755:107-120.

Birner P, Schindl M, Obermair A, Plank C, Breitenecker G, Oberhuber G. Overexpression of hypoxia-inducible factor 1alpha is a marker for an unfavorable prognosis in early-stage invasive cervical cancer. *Cancer Res*. 2000; 60:4693 - 4696.

Brahimi-Horn C, Mazure N, Pouyssegur J. Signalling via the hypoxia-inducible factor-1alpha requires multiple posttranslational modifications. *Cell Signal*. 2005; 17:1 - 9.

Bukowski R, Vaughn C, Bottomley R, Chen T. Phase II study of anguidine in gastrointestinal malignancies: a Southwest Oncology Group study. *Cancer Treat Rep*. 1982; 66:381 - 383.

Carmeliet P, Jain RK. Principles and mechanisms of vessel normalization for

cancer and other angiogenic diseases. *Nat Rev Drug Discov.* 2011; 10:417 - 427.

Chang CC, Lin MT, Lin BR, Jeng YM, Chen ST, Chu CY, Chen RJ, Chang KJ, Yang PC, Kuo ML. Effect of connective tissue growth factor on hypoxia-inducible factor 1alpha degradation and tumor angiogenesis. *J Natl Cancer Inst.* 2006; 19:984 - 995.

Cho YS, Kim CH, Park JW. Involvement of HIF-1alpha in UVB-induced epidermal hyperplasia. *Mol Cells.* 2009; 28:537 - 543.

Cole RJ, Cox RH. Handbook of Toxic Fungal Metabolites. The trichothecenes. Academic Press: New York, 1981. pp 152 - 263.

Dayan, F. Roux, D., Brahimi-Horn, MC, Pouyssegur, J, Mazure, NM. The oxygen sensor factor-inhibiting hypoxia-inducible factor-1 controls expression of distinct genes through the bifunctional transcriptional character of hypoxia-inducible factor-1 alpha. *Cancer Res.* 2006; 66:3688-3698.

Dawkins AW. Phytotoxic Compounds Produced by *Fusarium equiseti*, Part II. Chemistry of Diacetoxysirpenol. *J Chem Soc. Perkin Trans.* 1996; 1:116 - 123.

Dengler VL, Galbraith MD, Espinosa JM. Transcriptional regulation by hypoxia inducible factors. Crit Rev Biochem Mol Biol. 2014; 49:1 - 15.

Dosik GM, Barlogie B, Johnston DA, Murphy WK, Drewinko B. Lethal and cytotoxic effects of anguidine on a human colon cancer cell line. Cancer Res. 1978; 38:3304 - 3309.

Fandrey J, Gorr TA, Gassmann M. Regulation cellular oxygen sensing by hydroxylation. Cardiovasc Res. 2006, 71:642-651.

Ferrara N, Hillan KJ, Gerber HP, Novotny W. Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer. Nat Rev Drug Discov. 2004; 3:391 - 400.

Harris AL. Hypoxia - a key regulatory factor in tumour growth. Nat Rev Cancer. 2002, 2:38 - 47.

Hromas RA, Yung WK. Anguidine potentiates cis-platinum in human brain tumor cells. J Neurooncol. 1986; 3:343 - 348.

Huang D, Lan H, Liu F, Wang S, Chen X, Jin K, Mou X. Anti-angiogenesis or pro-angiogenesis for cancer treatment: focus on drug distribution. *Int J Clin Exp Med.* 2015; 8:8369 - 8376.

Hudson CC, Liu M, Chiang GG, Otterness DM, Loomis DC, Kaper F, Giaccia AJ, Abraham RT. Regulation of hypoxia-inducible factor 1alpha expression and function by the mammalian target of rapamycin. *Mol Cell Biol.* 2002; 22:7004 - 7014.

Isaacs JS, Jung YJ, Mimnaugh EG. Hsp90 regulates a von Hippel Lindau-independent hypoxia inducible factor-1 alpha-degradative pathway. *J Biol Chem.* 2002; 277:29936 - 29944.

Ivan M, Kondo K, Yang H, Kim W, Valiando J, Ohh M, Salic A, Asara JM, Lane WS, Kaelin WG Jr. HIF alpha targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ sensing. *Science.* 2001; 292:464 - 468.

Jaakkola P, Mole DR, Tian YM, Wilson MI, Gielbert J, Gaskell SJ, von Kriegsheim A, Hebestreit HF, Mukherji M, Schofield CJ, Maxwell PH, Pugh CW, Ratcliffe PJ. Targeting of HIF-alpha to the von Hippel-Lindau

ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science*. 2001; 292:468 - 472.

Jun DY, Kim JS, Park HS, Song WS, Bae YS, Kim YH. Cytotoxicity of diacetoxyscirpenol is associated with apoptosis by activation of caspase-8 and interruption of cell cycle progression by down-regulation of cdk4 and cyclin B1 in human Jurkat T cells. *Toxicol Appl Pharmacol*. 2007; 15:190 - 201.

Kong D, Park EJ, Stephen AG. Echinomycin, a smallmolecule inhibitor of hypoxia-inducible factor-1 DNA binding activity. *Cancer Res*. 2005; 65:9047 - 9055.

Lando D, Peet DJ, Gorman JJ, Whelan DA, Whitelaw ML, Bruick RK. FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev*. 2002; 16:1466-1471.

Lang KJ, Kappel A, Goodall GJ. Hypoxia-inducible factor-1 α mRNA contains an internal ribosome entry site that allows efficient translation during normoxia and hypoxia. *Mol Biol Cell*. 2002; 13:1792 - 1801.

Lee YM, Lim JH, Chun YS, Moon HE, Lee MK, Huang LE, Park JW. Nutlin-3,

an Hdm2 antagonist, inhibits tumor adaptation to hypoxia by stimulating the FIH-mediated inactivation of HIF-1 α . *Carcinogenesis*. 2009; 30:1768 - 1775.

Lee YM, Lim JH, Yoon H, Chun YS, Park JW. Antihepatoma activity of chaetocin due to deregulated splicing of hypoxia-inducible factor 1 α pre-mRNA in mice and in vitro. *Hepatology*. 2011; 53:171 - 180.

Lee SJ, No YR, Dang DT, Dang LH, Yang VW, Shim H, Yun CC. Regulation of hypoxia-inducible factor 1 α (HIF-1 α) by lysophosphatidic acid is dependent on interplay between p53 and Krüppel-like factor 5. *J Biol Chem*. 2013; 288:25244 - 25253.

Lou JJ, Chua YL, Chew EH, Gao J, Bushell M, Hagen T. Inhibition of hypoxia-inducible factor-1 α (HIF-1 α) protein synthesis by DNA damage inducing agents. *PLoS One*. 2010; 7:5(5).

Mahon PC, Hirota, K, and semenza, GL. FIH-1 : a novel protein that interacts with HIF-1 α and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev*. 2001; 15:2675-2686.

Ma J, Waxman DJ. Combination of antiangiogenesis with chemotherapy for more effective cancer treatment. *Mol Cancer Ther.* 2008; 7:3670 - 3684.

Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, Wykoff CC, Pugh CW, Maher ER, Ratcliffe PJ. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature.* 1999; 399:271 - 275.

Mills CN, Joshi SS, Niles RM. Expression and function of hypoxia inducible factor-1 alpha in human melanoma under non-hypoxic conditions. *Mol Cancer.* 2009; 8:104.

Press MF, Lenz HJ. EGFR, HER2 and VEGF pathways: validated targets for cancer treatment. *Drugs.* 2007; 67:2045 - 2075.

Pugh CW, O'Rourke JF, Nagao M, Gleadle JM, Ratcliffe PJ. Activation of hypoxia-inducible factor-1; definition of regulatory domains within the alpha subunit. *J Biol Chem.* 1997, 272:11205-11214.

Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: role of the HIF

system. Nat Med. 2003, 9:677 - 684.

Ryan HE, Poloni M, McNulty W, Elson D, Gassmann M, Arbeit JM, Johnson RS. Hypoxia-inducible factor-1alpha is a positive factor in solid tumor growth. Cancer Res. 2000; 60:4010 - 4015.

Rapisarda A, Uranchimeg B, Sordet O. Topoisomerase I-mediated inhibition of hypoxia-inducible factor 1: mechanism and therapeutic implications. Cancer Res. 2004; 64:1475 - 1482.

Ricker JL, Chen Z, Yang XP, Pribluda VS, Swartz GM, Van Waes C. 2-methoxyestradiol inhibits hypoxia-inducible factor 1alpha, tumor growth, and angiogenesis and augments paclitaxel efficacy in head and neck squamous cell carcinoma. Clin Cancer Res. 2004; 15:8665 - 8673.

Rocha, S. Gene regulation under low oxygen :holding your breath for transcription. Trends Biochem Sci. 2007, 32:389-397.

Semenza GL. HIF-1: mediator of physiological and pathophysiological responses to hypoxia. J Appl Physiol. 1985; 88:1474 - 1480.

Shin DH, Chun YS, Lee DS, Huang LE, Park JW. Bortezomib inhibits tumor adaptation to hypoxia by stimulating the FIH-mediated repression of hypoxia-inducible factor-1. *Blood*. 2008; 15:3131 - 3136.

van Egmond HP, Schothorst RC, Jonker MA. Regulations relating to mycotoxins in food: perspectives in a global and European context. *Anal Bioanal Chem*. 2007; 389:147 - 157.

Wang GL, Semenza GL. Purification and characterization of hypoxia inducible factor 1. *J Biol Chem*. 1995, 270:1230 - 1237.

Yap HY, Murphy WK, DiStefano A, Blumenschein GR, Bodey GP. Phase II study of anguidine in advanced breast cancer. *Cancer Treat Rep*. 1979; 63:789 - 791.

Yeo EJ, Chun YS, Cho YS, Kim J, Lee JC, Kim MS, Park JW. YC-1: a potential anticancer drug targeting hypoxia-inducible factor 1. *J Natl Cancer Inst*. 2003; 2:516 - 525.

Zhang H, Qian DZ, Tan YS. Digoxin and other cardiac glycosides inhibit

HIF-1 α synthesis and block tumor growth. Proc Natl Acad Sci USA. 2008; 105:19579 - 19586.

Zhong H, Chiles K, Feldser D, Laughner E, Hanrahan C, Georgescu MM, Simons JW, Semenza GL. Modulation of hypoxia-inducible factor 1 α expression by the epidermal growth factor/phosphatidylinositol 3-kinase/PTEN/AKT/FRAP pathway in human prostate cancer cells: implications for tumor angiogenesis and therapeutics. Cancer Res. 2000; 15:60(6):1541-5.

국 문 초 록

저산소 유도인자 (HIF)는 세포의 저산소 적응과정에서 핵심적인 역할을 한다. 특히 고형암은 저산소 상황에서 암세포의 생존과 혈관형성에 필요한 HIF를 다량으로 발현하므로 HIF를 표적으로 하는 치료제를 개발하는 것이 중요하다. 본 연구에서는 HIF를 효과적으로 억제할 수 있는 새로운 약물을 찾기 위한 실험을 진행하였다. 총 7,280개의 화합물 후보군은 EPO 프로모터와 Luciferase 퓨전 벡터를 리포터로 넣은 간암세포주인 Hep3B를 이용하여 HIF의 활성도를 측정하였으며 그 중에서 최종 후보군을 선정하였다. 구조분석을 통해 최종후보군으로 선택된 단일물질이 diacetoxyscirpenol임을 확인하였다. Diacetoxyscirpenol은 여러 암세포에서 HIF의 단백질 발현을 억제할 뿐만 아니라, HIF의 활성 및 HIF에 의해 유도되는 하위유전자들의 발현을 억제하였다. 또한 혈관형성과 암세포의 성장을 억제하며 동물실험에서도 암의 성장을 효과적으로 억제하였다. Diacetoxyscirpenol은 HIF의 합성과정을 억제하며 HIF와 ARNT의 결합을 억제함으로써 저산소 관련인자의 발현을 억제함을 확인하였다. 이러한 결과를 통해 본 연구는 Diacetoxyscirpenol가 저산소 조건에 노출되어 있는 고형암을 치료할 수 있는 새로운 약물로서 사용될 수 있을 것으로 사료된다.

주요어 : 암, 저산소, HIF-1, Diacetoxyscirpenol, Drug screening.

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